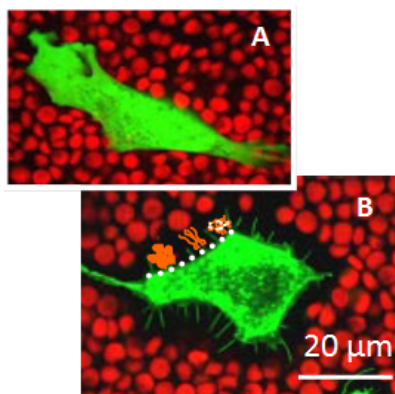




Biological Relevance of HA-Rich Matrices

Description

In tissues, cells are embedded in extracellular matrix (ECM) which is made from components that are produced and then secreted by the cells. This matrix has many functions : it serves as integrative support to anchor cells, it distinguishes tissues from one another, it regulates inter-cellular communication and it ensures the mechanical stability of the tissue Alberts et al., 2008. Many mammalian cells additionally endow themselves with a pericellular matrix (PCM), also referred to as a cellular coat or glycocalyx, which is located at the interface between the ECM and the cellular membrane, and typically defined by its direct anchorage to the plasma membrane Evanko et al., 2007. Pericellular matrices have been reported for a variety of cells *in vivo* and in tissue culture such as fibroblasts Hedman et al., 1979, chondrocytes Evanko et al., 2007 Knudson, 1993, epithelial cells Cohen et al., 2003, prostate cells Ricciardelli et al., 2007, monocytes Sabri et al., 2000 and endothelial cells Smith et al., 2003 Constantinescu et al., 2003. The thickness of the coat varies from one cell type to another and depends strongly on the state of the cell (Figure 1) Kultti et al., 2006 McLane et al., 2013. For example, monocyte coats typically extend outwards by only 10 to 20 nm Sabri et al., 2000, whereas 20 μm thick coats were observed on chondrocytes Evanko et al., 2007 Claris & Fraser, 1968.

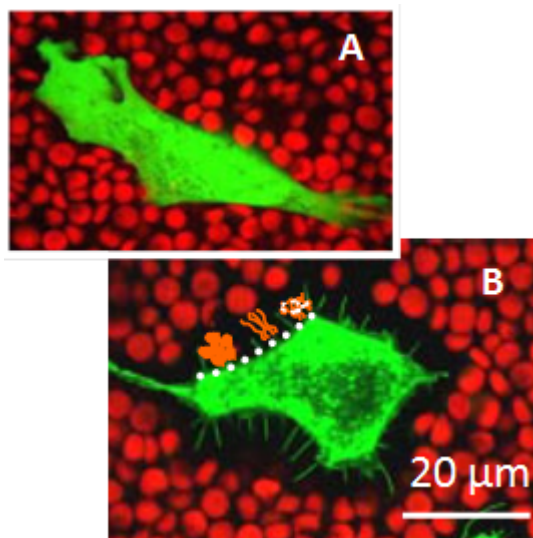


Figure 1. Visualization of a PCM by a particle exclusion assay. Red blood cells (red) were settled on a culture of human breast adenocarcinoma cell line, MCF-7, and pericellular matrices become visible because they cannot be penetrated by red blood cells. Membranes of MCF-7 cells were stained with a fluorescent dye (green) A. Native MCF-7 cell. B. MCF-7 cell transfected to overproduce hyaluronan (HA); membrane protrusions, so called microvilli (visible as green lines) were induced by transfection. Schematics (orange) were drawn into the micrograph to illustrate that the thick coat can be the result of different HA configurations: a cross-linked multilayer, extended chains in a brush-like conformation, or extended chains decorated with additional proteins (from left to right).

Copyright: Adapted from Kultti, A., Rilla, K., Tiihonen, R., Spicer, A. P., Tammi, R. H., and Tammi, M. I. Hyaluronan synthesis induces microvillus-like cell surface protrusions. *The Journal of biological chemistry* 281, 15821-15828 (2006)

For a long time, the PCM was considered as a passive lubricating layer around the cells, because of its strongly hydrated gel-like nature. Later, it was found that it also plays an active role in many biological processes Toole, 2001, including cell adhesion Cohen et al., 2006 Siegelma, et al., 1999, cell proliferation Evanko et al., 1999 Brecht et al., 1986, cell migration Ricciardelli et al., 2007 Itano et al., 2002, the display of growth factors Macri et al., 2007, embryogenesis Toole, 2001 and fertilization Russell & Salustri, 2006. Although the PCM is involved in a diverse range of fundamental biological processes the relation between its structure and functions remains poorly understood.

A key structural component of many pericellular matrices is the polysaccharide hyaluronan. It is synthesized directly at the cellular membrane and can remain anchored to the cell surface either at the HA-synthase that produced the HA chain or it can bind to the cell surface through HA-binding receptors. The linear HA chains of typically several micrometer in contour length alone are not enough to assemble PCMs of several microns in thickness Wolny et al.,. Instead, they serve as a scaffold or polyvalent template Day & Sheehan, 2001 for the incorporation of members of the family of hyaluronan-binding proteins, also called hyaladherins. For example, the attachment of aggrecan, a proteoglycan that has the form of a bottle brush, to HA chains results in the formation of extended and highly hydrated aggregated Knudson et al., 1993 that can give rise to thick cellular coats.

Variations in the length of HA chains, the confinement of HA to the cell surface, and the non-uniform distribution of various hyaladherins along one chain can give rise to a spatial organization of the PCM. In cultures of rat chondrocytes, for example, the density of the coat was shown to be relatively high at a distance of about 1 μm from the cell membrane, with a mesh size of 100 nm, as compared to the periphery, where the mesh size reached 500 nm McLane et al., 2013. Such density gradients in the PCM create an osmotic force which decays with the distance from the cell surface McLane et al., 2014. Also, when cells were transfected with HA synthases, the thickness of the HA-rich coat was found to increase drastically, driven by the formation of membrane protrusions (microvilli ; Figure.1). Interestingly, the increase in the density of HA on the cellular membrane was proposed as the driving force for microvillus formation Kultt et al., 2006 iRilla et al., 2008.

PCMs are highly dynamic structures Lee et al., 1993, they reorganize constantly as a function of external stimuli. The dynamic rearrangement of the coat can be observed when cells detach from the surface or from surrounding cells, like during mitosis or migration Brecht et al., 1986 Horwitz & Parsons, 1999. Detachment of the cells in these cases is characterized by a swollen PCM and a rounded cell

shapeBrecht et al., 1986.

Under inflammation, for example in arthritis, the PCM undergoes remodeling – the HA matrix needs to be cross-linked to stabilize the entire assembly and to prevent the loss of tissue. Day & de la Motte, 2005. Remodeling of the PCM structure by HA cross-linking was proposed as a pathway to regulate inflammatory process. For example, the formation of cable-like structures, larger than 200 microns in length, with altered leukocyte-binding properties, were observed in response to inflammatory stimuli de la Motte et al., 2003. These cable-like structures were suggested to be formed by cross-linked HA chains. Day & de la Motte, 2005. The mechanisms of PCM cross-linking as well as how cross-linking alters the physico-chemical properties of the HA matrix and its functional activity remain poorly understood. Various proteins and protein complexes were proposed to cross-link HA meshworks (Figure 2).

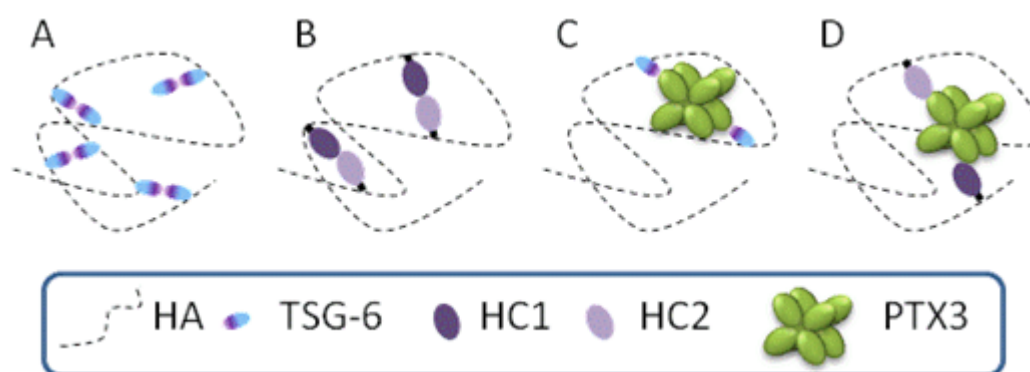


Figure.2 Hypothesized mechanisms of TSG-6 mediated HA cross-linking. A. Direct cross-linking of HA chains through self-association between TSG-6 monomers. B. HA cross-linking via non-covalent interaction between heavy chains (HCs) of inter- α -inhibitor ($I\alpha I$), covalently transferred onto HA by TSG-6. C. A cross-linking node formed by a complex of TSG-6 and PTX3. PTX3 is a multimeric protein, which does not bind HA directly but that interacts with TSG-6 and $I\alpha I$. D. Incorporation of PTX3 cross-linker can be mediated via HCs of $I\alpha I$.

Several different cross-linking pathways might be at play simultaneously in a given tissue. Considering the highly dynamic nature of HA and the PCM, it is likely that HA-rich matrices are typically stabilized by transient cross-linking interactions, allowing for continuous matrix remodeling and a constantly changing network of interactions.

Category

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