

Protein X-ray crystallography and glycobiology

Dr Annabelle Varrot

« Structural Glycosciences » Summer School

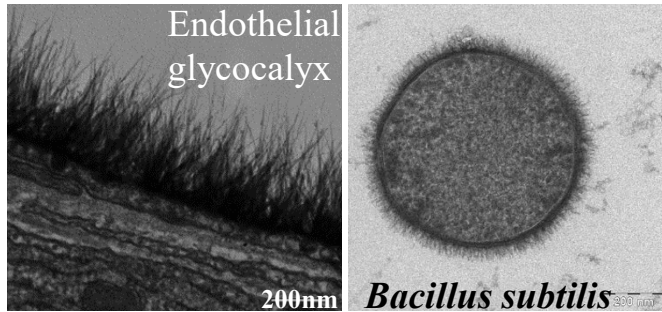
IBS, Grenoble 7th June 2023



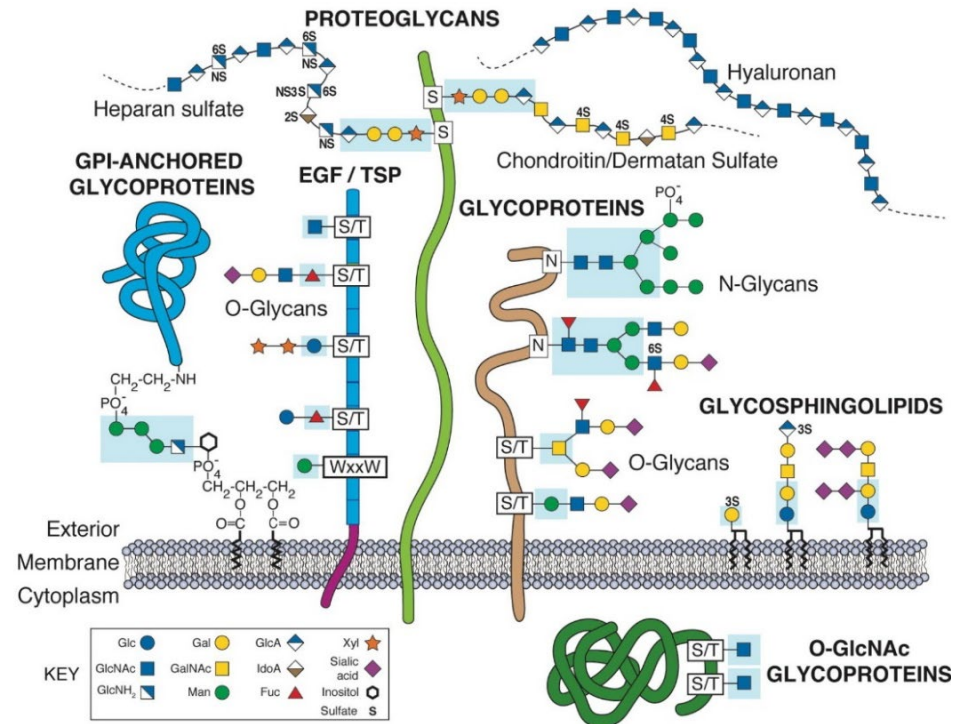
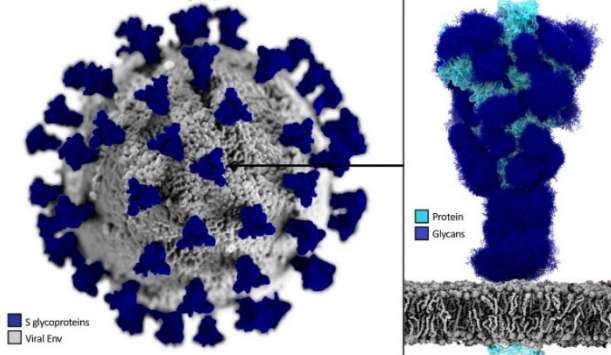
"Sweet side" of cells



- Sugars: 3rd alphabet of life: Glycome → glycode
- Every cell is covered by **glycoconjugates**



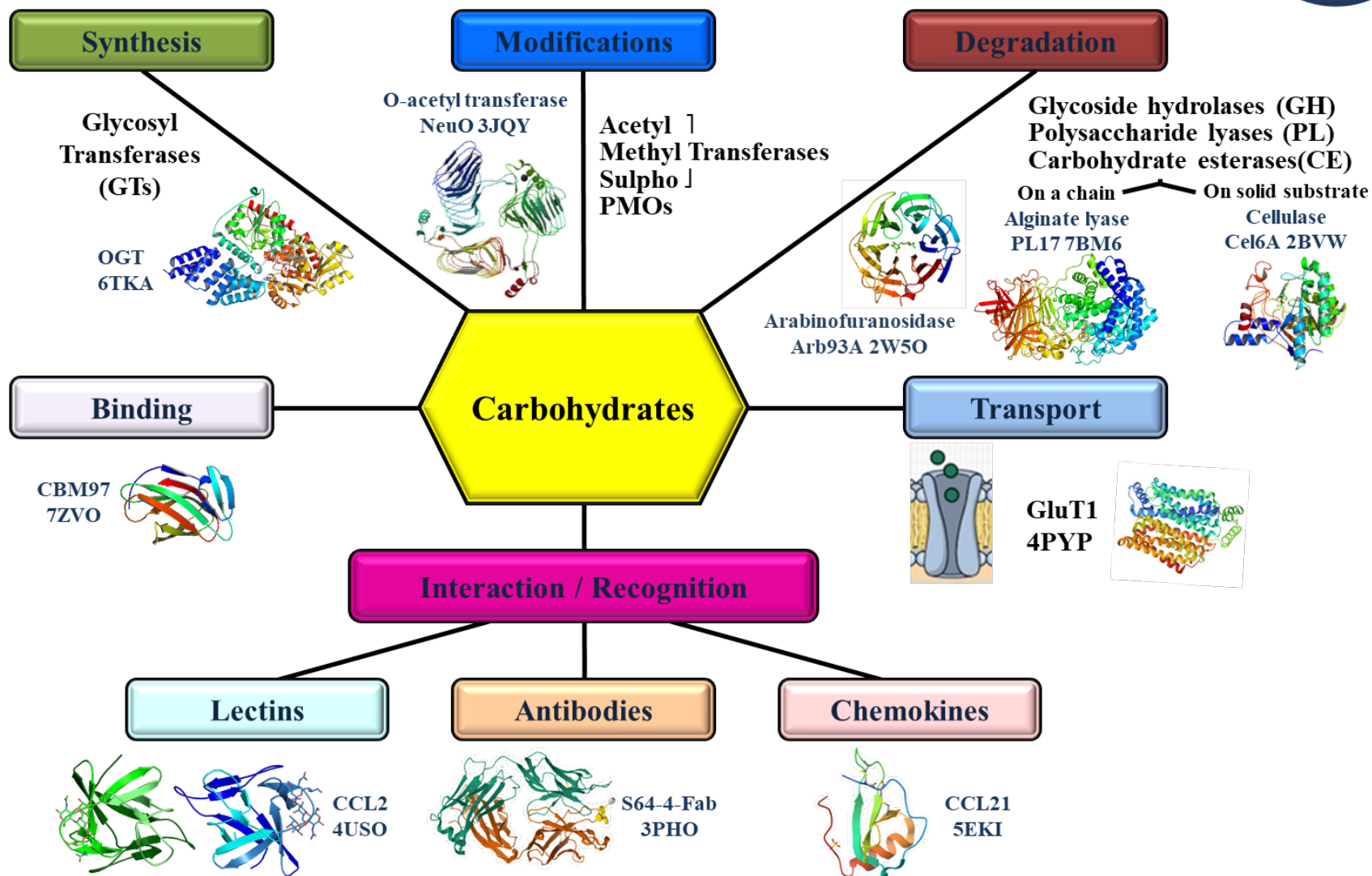
SARS-CoV-2, glycan shield



- Essentials in cell identity, fate, recognition & signaling

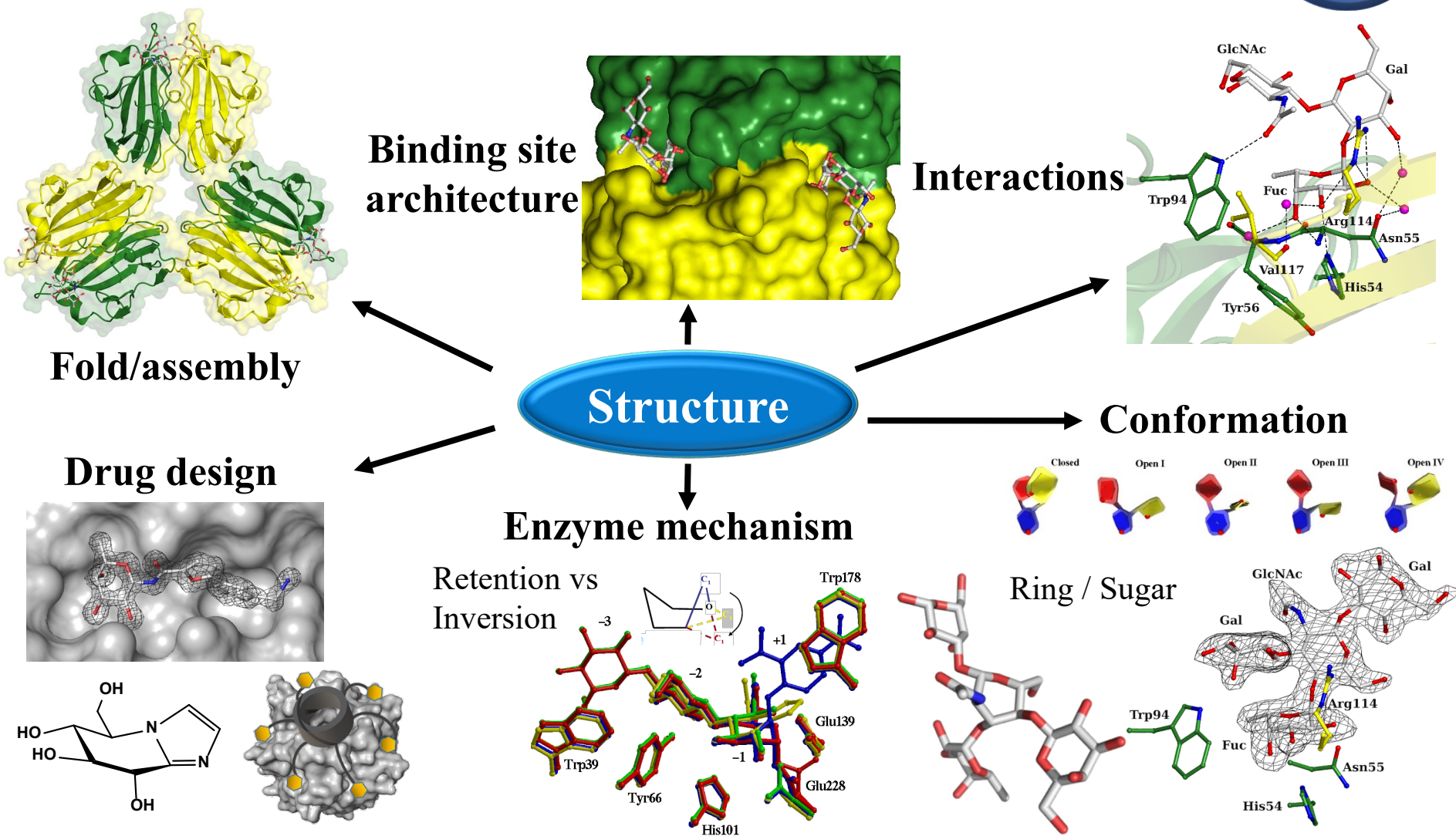
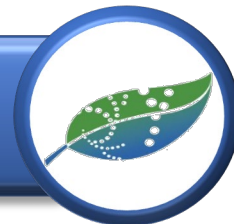


Protein-carbohydrate interactions

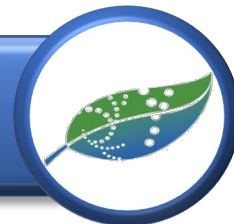




Which info by X-ray crystallography?



Protein X-ray crystallography

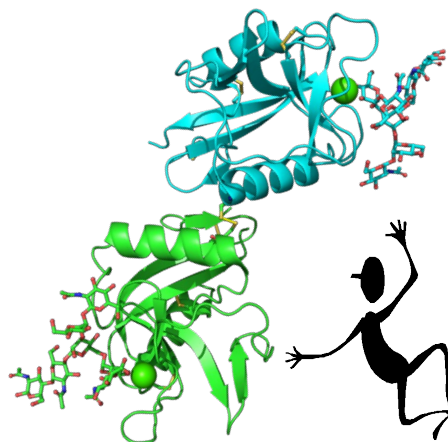


➤ Advantages

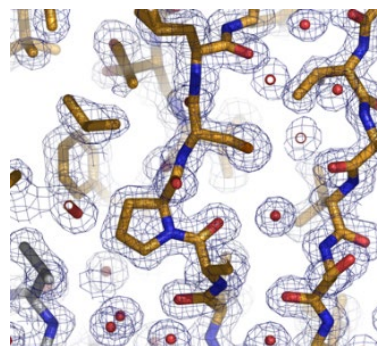
- Can go to atomic resolution
- Atomic details obtained

➤ Disadvantages

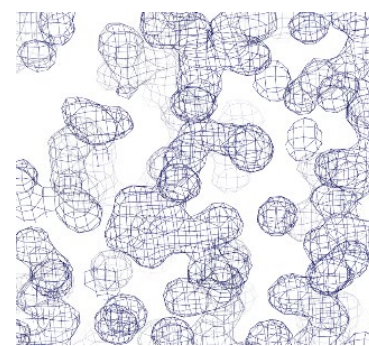
- Molecules in solid-state environment
- Require crystals
- Requires order to diffract



Refinement



Modelling



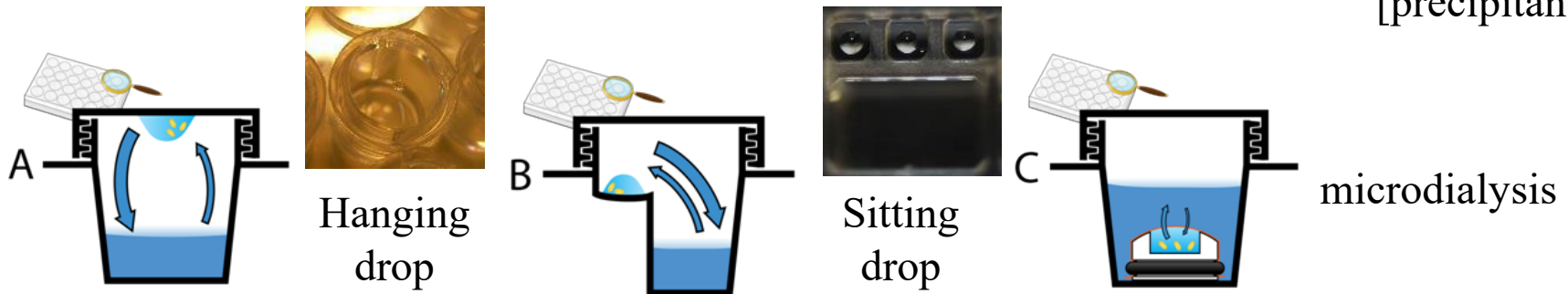
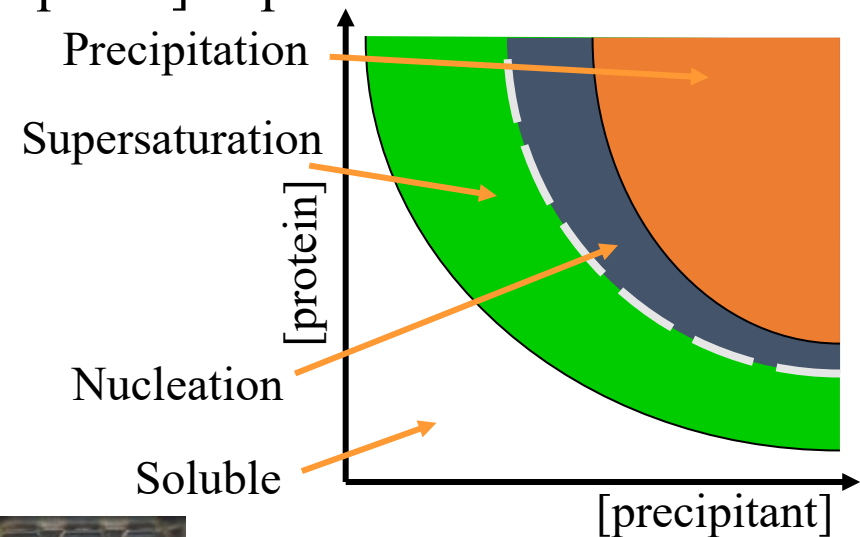
Phasing



Protein crystallogenesi



- Need pure and monodisperse protein
- Empirical
 - pH, buffer, temperature, [salt], [precipitant] dependant
- Precipitants
 - High salt concentration
 - Alcohols or volatil compounds
 - Organic polymers
- Manual or robotized
- Vapor diffusion mostly used



How to obtain crystals of protein complexes with ligand

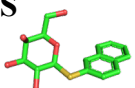
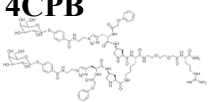

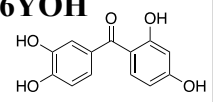
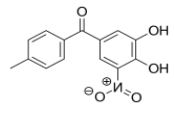



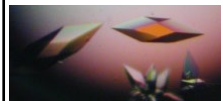

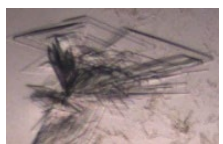

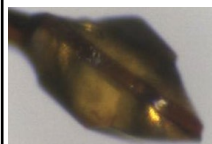
Cocrystallization



- Put the ligand in reservoir
- Preincubate with the protein
 - Limit protein dilution: $>1/10$
 - Set duration and temperature
 - Easily reproducible & require little ligand



- Can have crystal and spacegroup when change ligand: LecA

Gal α 1-2Gal β 2WYF	Gal α 1-3Gal β 1- 4Glc / 2VXJ	Gal α 1-6Glc / 4AL9	DEG144 / 4A6S 	Bivalent / 4CPB 	Cathecol 6YO3 	Cathecol 6YOH 	Tolcapone 
20% PEG6K 1 M LiCl 0.1 M NaAc 4	10% PEG5Kmm 25 mM KSCN 0.1 M NaAc 4.6	20% PEG2Kmm 0.2 M KBr 0.1 M NaAc 4.6	0.8 M Li ₂ SO ₄ 0.1 M NaAc 4.5	10 % PEG8K 10% PEG1K 0.2 M MgCl ₂ 0.1 M Tris 8.5	20% PEG6K 1 M LiCl 0.1 M NaAc 4.5 1% DMSO	20% PEG6K 1 M LiCl 0.1 M NaAc 4.5 1% DMSO	24% PEG2Kmm 0.1M KSCN 0.1M NaAc 4.5
2.4 Å, P2 ₁	1.9 Å, P1 _{79.2 86.5} 119.1 93.9 98.2 90.1	1.75 Å, P1 _{50.1} 58.1 75.9 101.1 92.9 101	2.15 Å, P3 ₂ 21	2.15 Å, P2 ₁ 2 ₁ 2 ₁	1.84 Å, I2	1.84 Å, P2 ₁ 2 ₂ 1	1.3 Å, C222 ₁
							

Soaking



➤ Dry soaking

- Let 0.1-1 μL ligand solution to dry before making drop

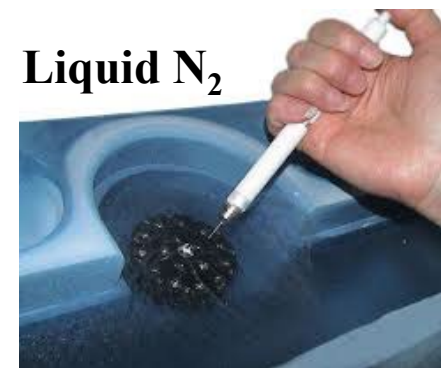
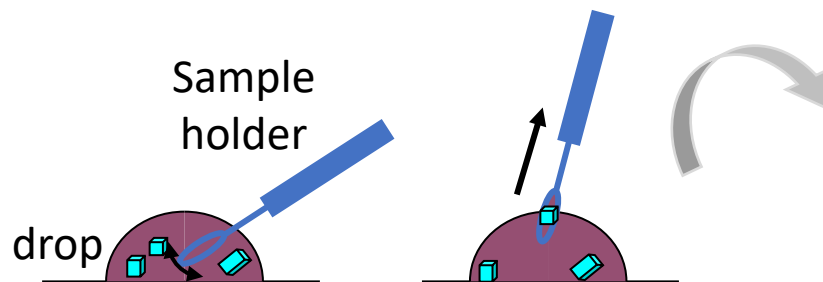
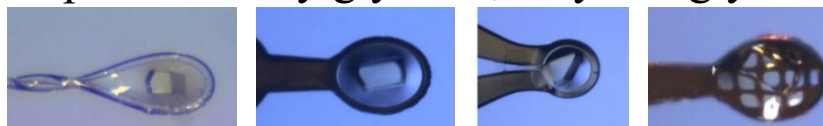
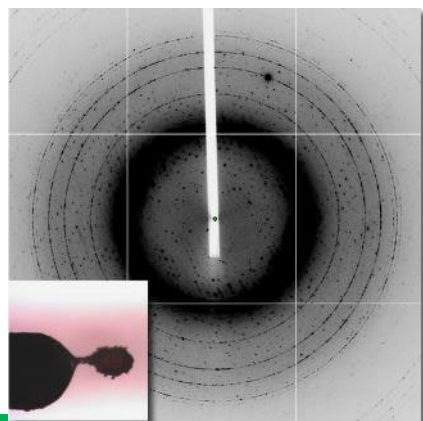
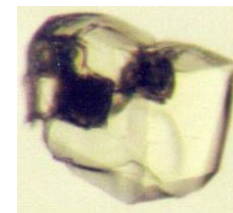
➤ Add the ligand to the reservoir solution

- Add concentrated solution or a bit of powder
- Transfer the crystal in solution with ligand
 - Try different concentrations and soaking time



➤ Soaking whilst freezing

- Freezing limits radiation damages
- Add ligand to the cryoprotectant solution
 - Less chance of replacement by glycerol, ethylene glycol and MPD

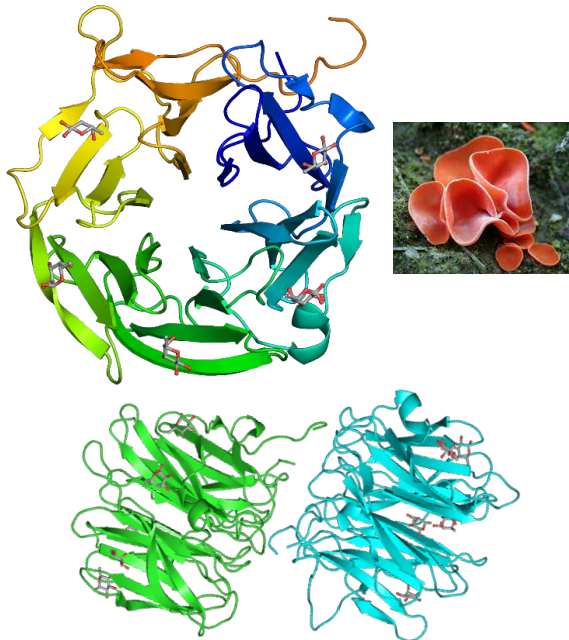


Phasing: molecular replacement

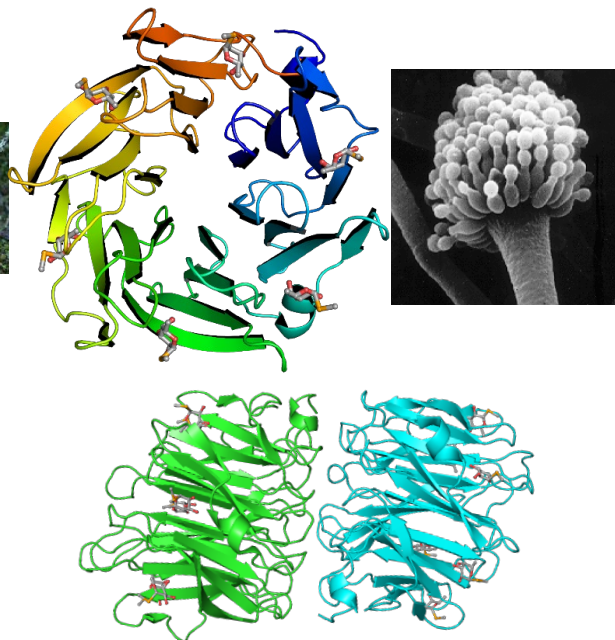


- Have a good model from homologous protein
 - Sequence identity > 25%
 - Can use alphafold if in Uniprot or fold known
 - Good conservation secondary structure: difficult for β -sheets

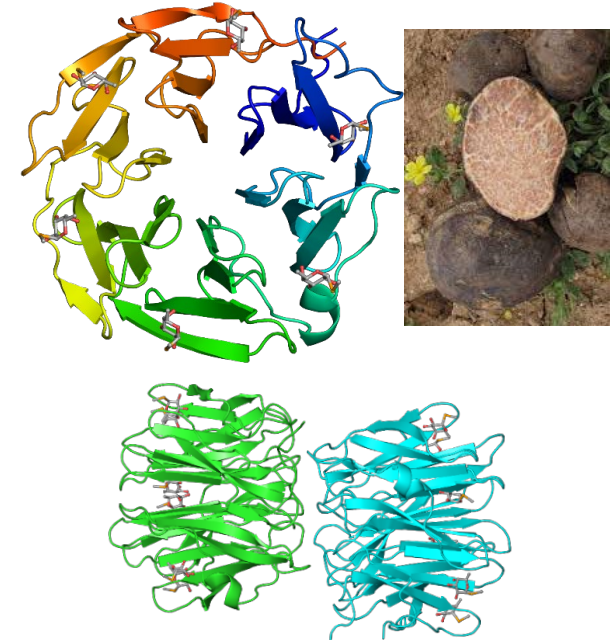
AAL/1OFZ/Hg
Aleuria auratia



FleA/4AGI/SFU (35%)
Aspergillus fumigatus

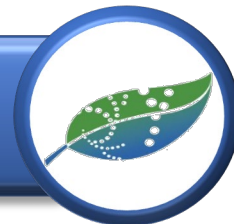


TrfbL1/SFU (32/43%)
Terfezia boudieri





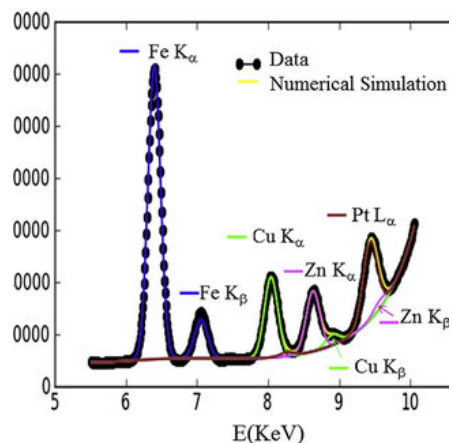
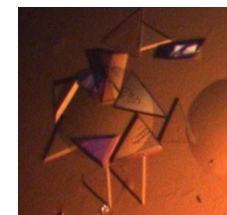
Phasing: isomorphous or anomalous



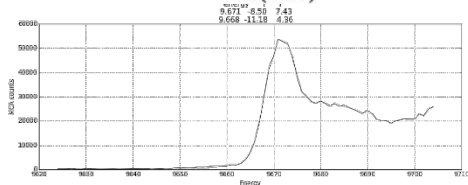
➤ Look in your crystallization conditions

■ PhoSL from *Pholiota squarrosa*

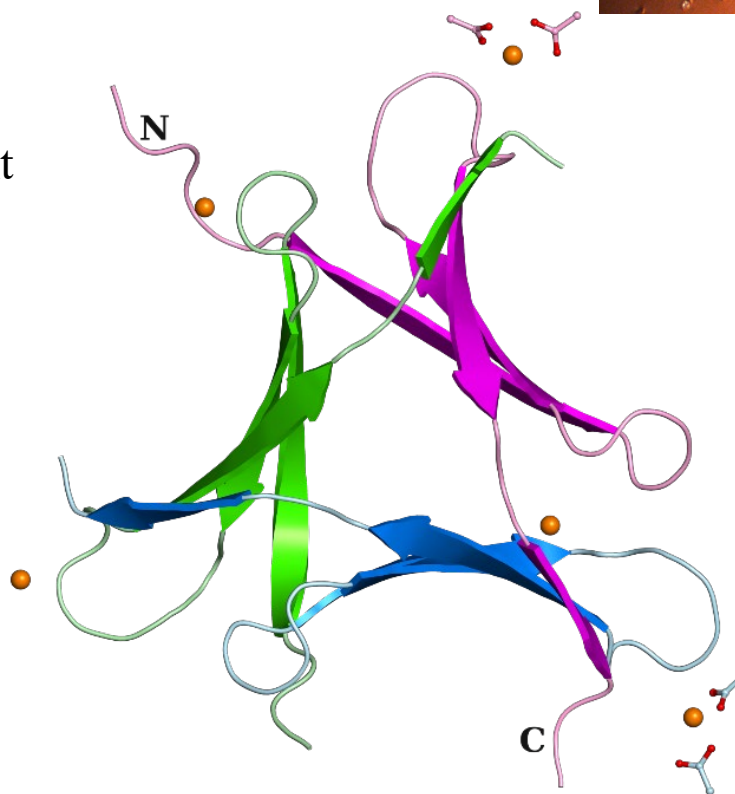
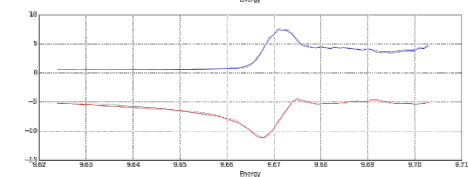
○ 300 mM Zinc acetate, 0.1 M Imidazole-HCl pH 6-7



X-ray fluorescent
XRF scan



Absorption
scan

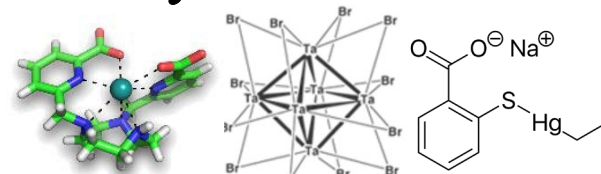




Phasing: isomorphous or anomalous-2

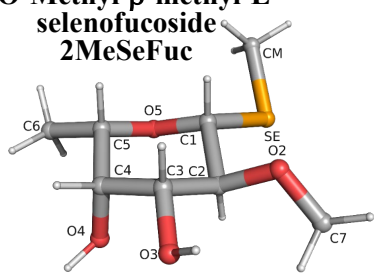


- Modified your protein with selenomet or selenocys
- Use heavy metals, clusters of lanthanides
- Use ligand with heavy atoms (Se, Br, S, F)



Lb-Tec2

2-O-Methyl-β-methyl-L-selenofucoside
2MeSeFuc



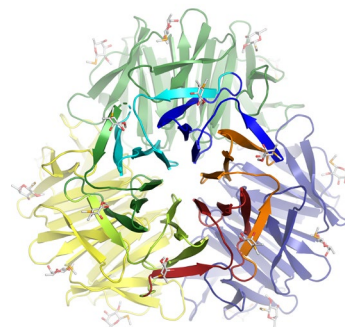
PolyAla building



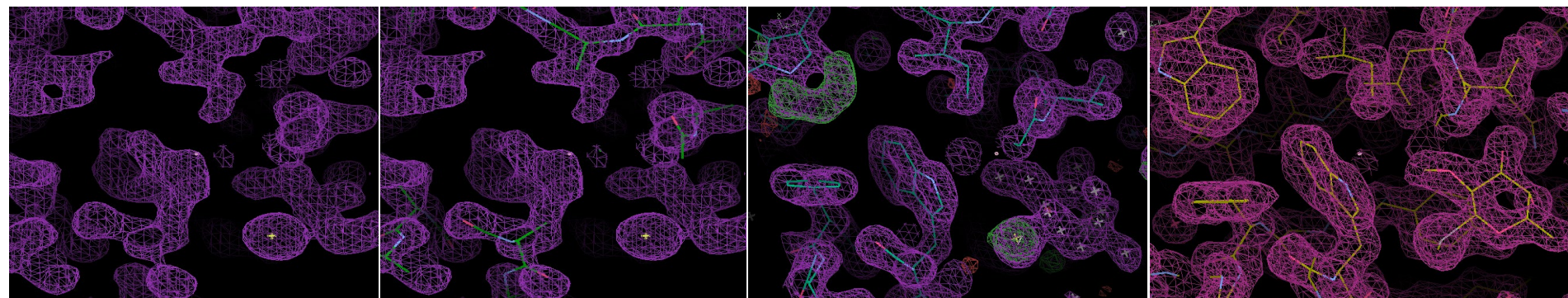
Alex Titz
HIPS
Saarbrücken
Germany
R Sommer



Sequence assigned



Final

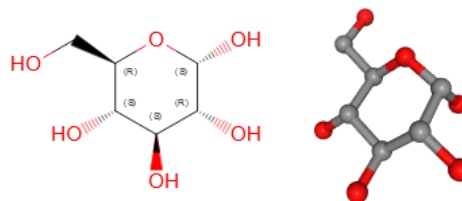


Carbohydrate refinement



➤ Find proper 3-letter code

- Check ligand database <http://ligand-expo.rcsb.org/ld-search.html>



GLC

alpha-D-glucopyranose

Find entries where: GLC

- is present as a standalone ligand in 301 entries
- as a non-polymer is covalently linked to polymer or other heterogen groups 61 entries
- is present in a branched oligosaccharide 1,638 entries

Ligand Expo Search Result Summary

Query: glucopyranose

Query type: Molecular name (exact sub-string)

Result count: 252

- 1 code per anomer

- α -D-Glucose: GLC
- β -D-Glucose: BGC
- β -L-Glucose: Z8T



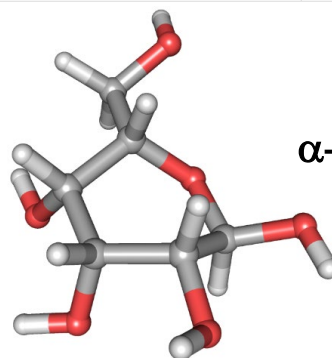
- Check library

- No distortion

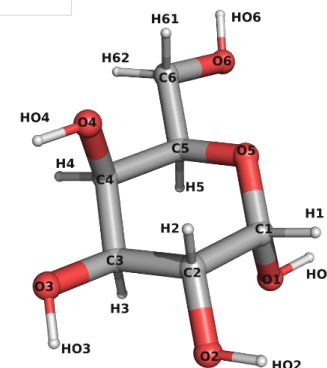
Chemical Component Summary	
Name	alpha-D-glucopyranose
Synonyms	alpha-D-glucose; D-glucose; glucose
Identifiers	(2S,3R,4S,5S,6R)-6-(hydroxymethyl)oxane-2,3,4,5-tetrol
Formula	C ₆ H ₁₂ O ₆
Molecular Weight	180.16
Type	D-SACCHARIDE, ALPHA LINKING
Isomeric SMILES	C([C@@H]1[C@H]([C@@H]([C@H]([C@@H](O1)O)O)O)O)O
InChI	InChI=1S/C6H12O6/c7-1-2-3(8)4(9)5(10)6(11)12-2/h2-11H,1H2/t2-,3-,4-,5-,6-/m1/s1
InChIKey	WQZGKKKJUFFOK-DVKNGEFBSA-N

Chemical Details

Formal Charge	0
Atom Count	24
Chiral Atom Count	5
Bond Count	24
Aromatic Bond Count	0



GLA
 α -D-galactose

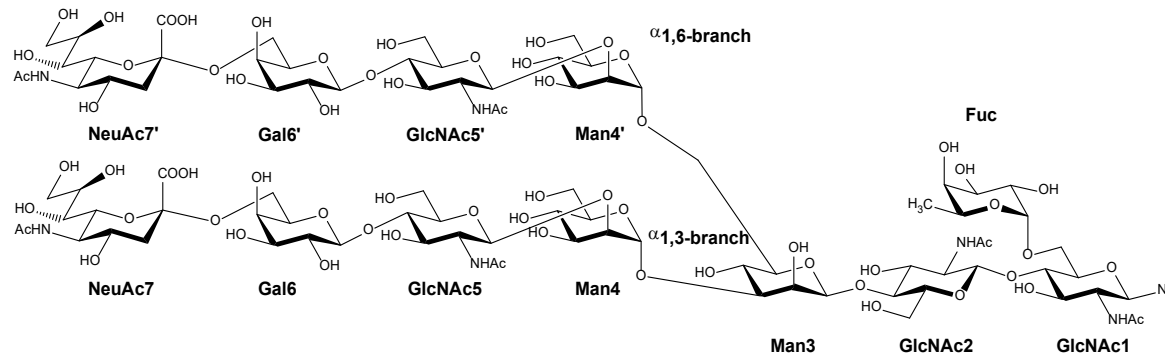


Oligosaccharide refinement

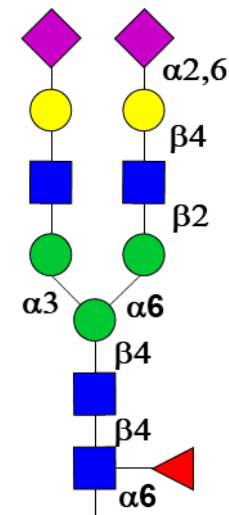


➤ Define correct linkage description

- Programs do not know how to deal with L-sugars



6FX3

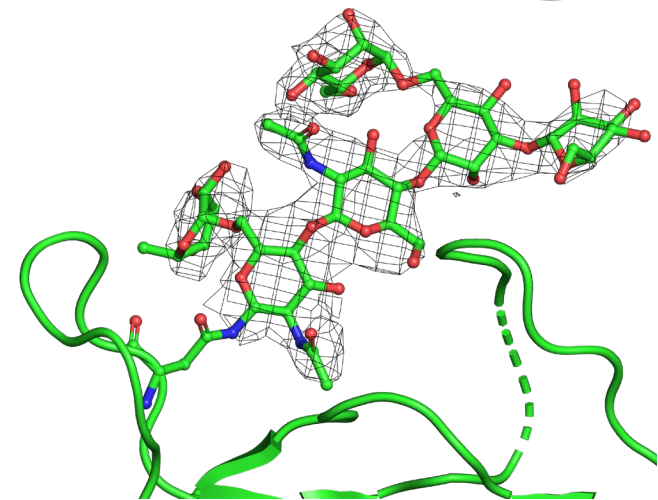
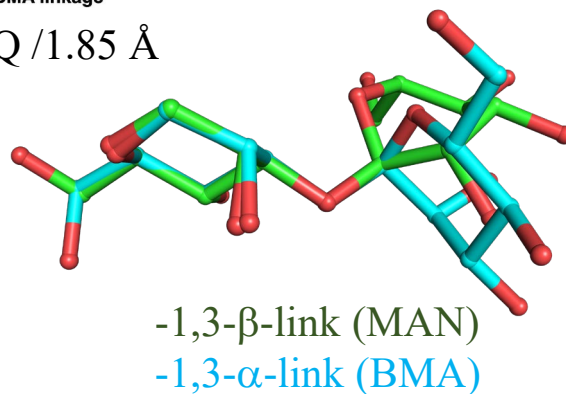
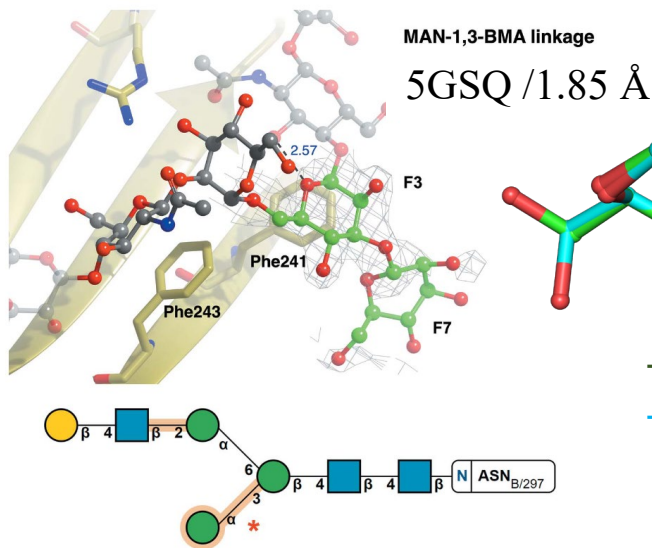


LINKR	O6	C4W	A	102	C1	FUC	A	101	BETA1-6
LINKR	O4	C4W	A	102	C1	NAG	A	103	BETA1-4
LINKR	O4	NAG	A	103	C1	BMA	A	104	BETA1-4
LINKR	O6	BMA	A	104	C1	MAN	A	105	ALPHA1-6
LINKR	O2	MAN	A	105	C1	NAG	A	106	BETA1-2
LINKR	O4	NAG	A	106	C1	GAL	A	107	BETA1-4
LINKR	C2	SIA	A	108	O6	GAL	A	107	SIA-GAL
LINKR	O3	BMA	A	104	C1	MAN	A	109	ALPHA1-3
LINKR	O2	MAN	A	109	C1	NAG	A	110	BETA1-2

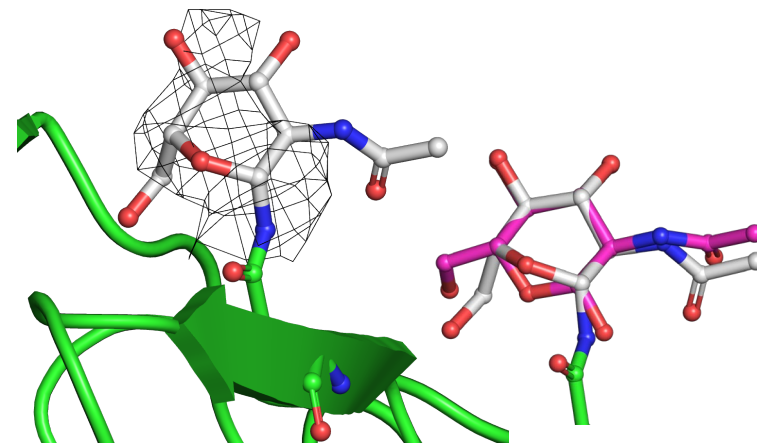
Carbohydrate validation



- Do not overfit at low resolution
- Check nomenclature, ring conformation, density fit
 - Privateer
 - PDB-REDO



7ALK /3 Å

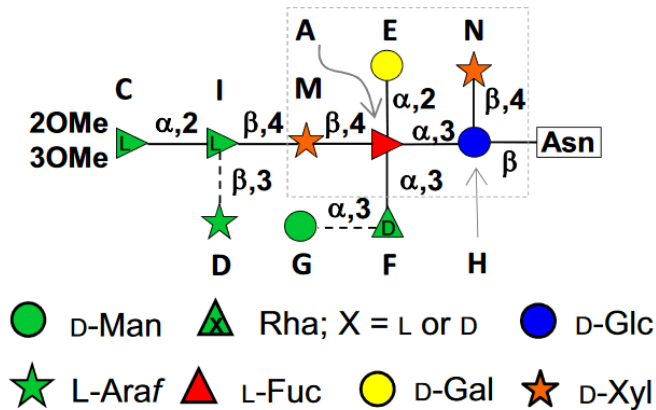




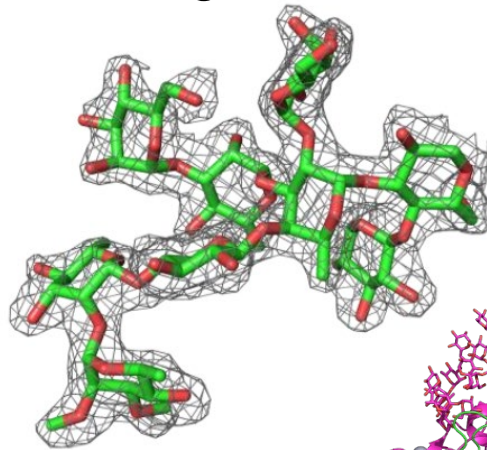
Tricky for non structural glycobioologists



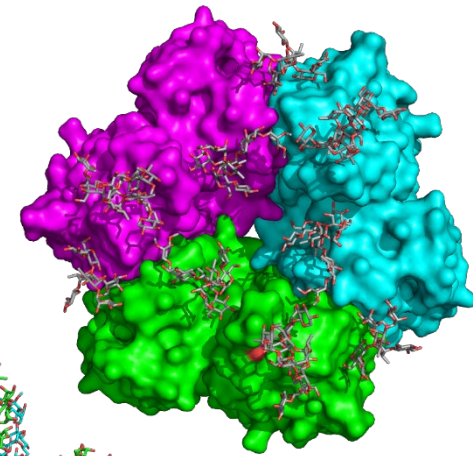
- Major capsid protein (Vp54) of chlorovirus PBCV-1
 - X-ray 2002: try to fit classical N-glycans
 - Sugar NMR 2013: highly complex N-glycosylation
 - X-ray revised in 2017 + modelling



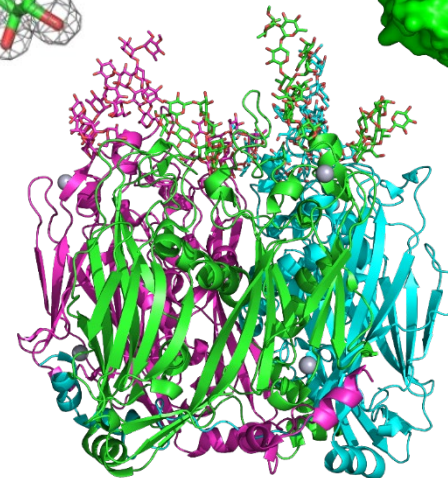
H = BGC A = FUC
 N/M = XYP E = GLA
 F = XXR G = MAN
 I = RM4 C = 7CV



5TIP

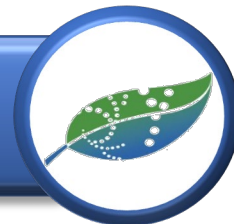


4 glycosylation sites



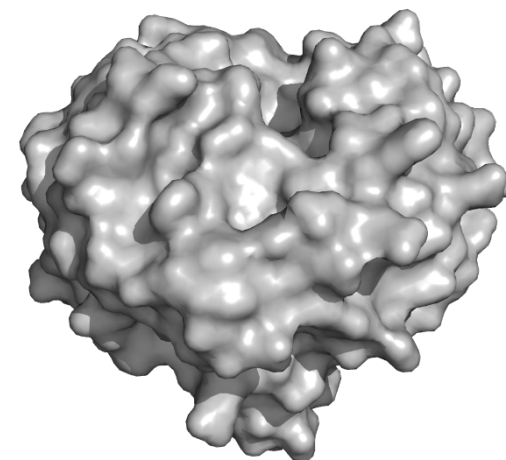
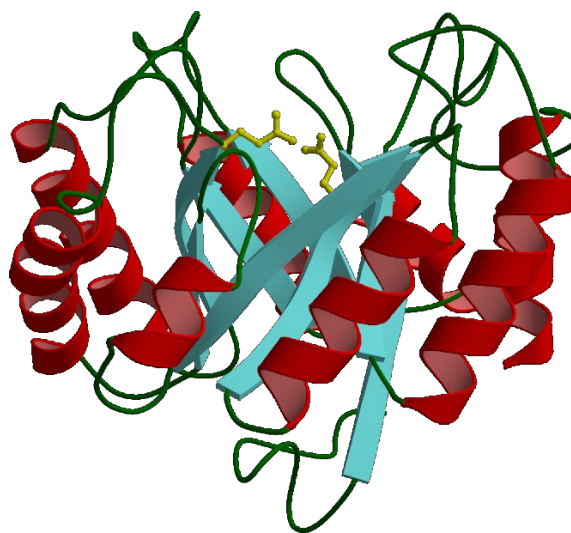
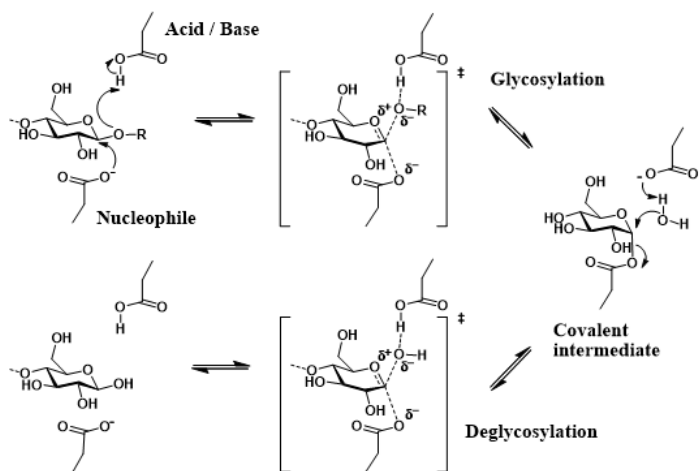
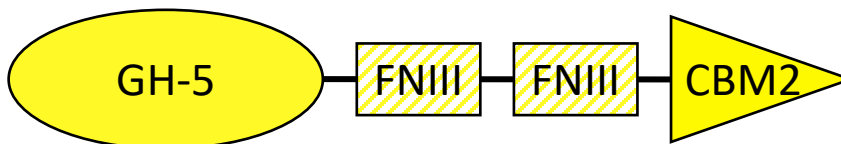


Insights in the retaining mechanism of GHs



➤ Cellulase Cel5A from *Bacillus agaradhaerens*

- Modular endoglucanase
- Active pH range 5.0-13.0
- Glu139 (acid/base) and Glu228 (nuc)

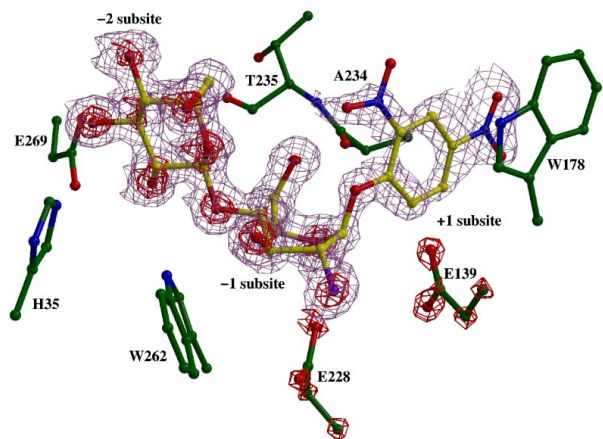
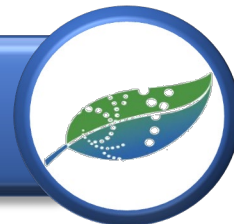


➤ Trapping of each step by X-ray crystallography

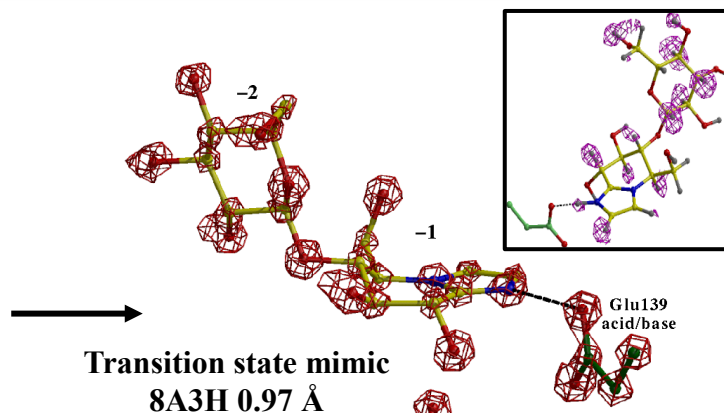
- Use specific ligand, mutated protein, inactive pH



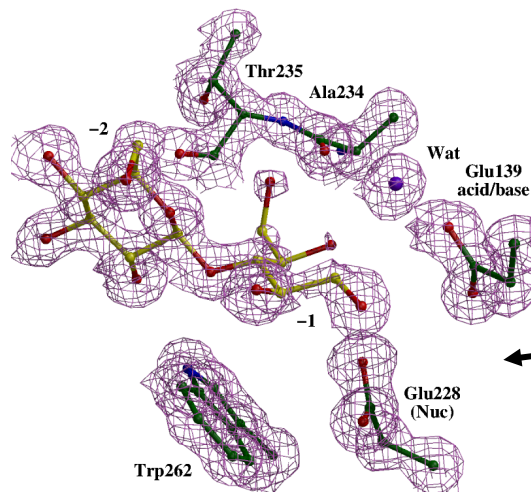
Snapshots at atomic resolution



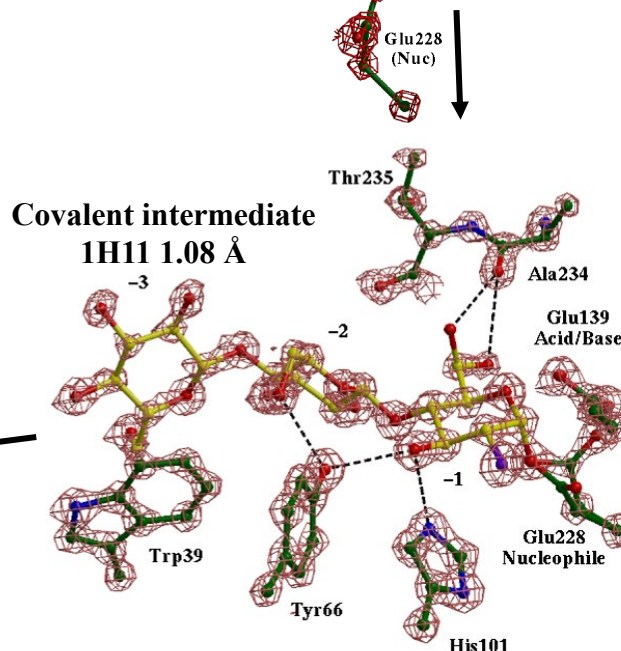
Mickaelis complex 1H2J 1.15 Å



Transition state mimic
8A3H 0.97 Å

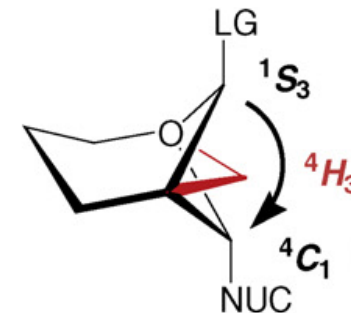
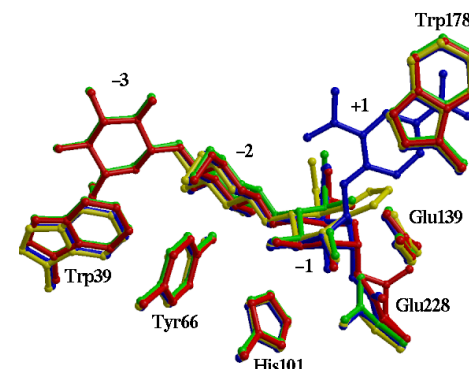


Product complex 1HF6 1.15 Å



Covalent intermediate
1H11 1.08 Å

- Static Cel5A
- Nucleophilic migration



GHs inhibitors design

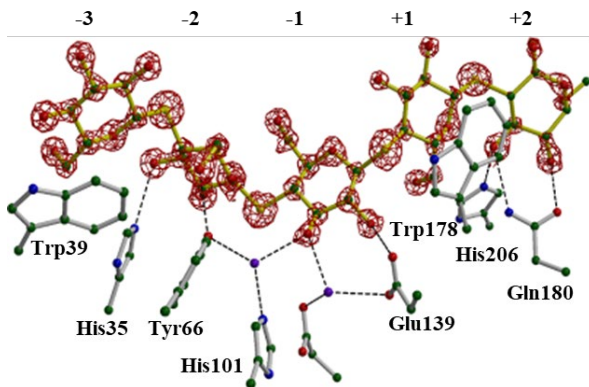
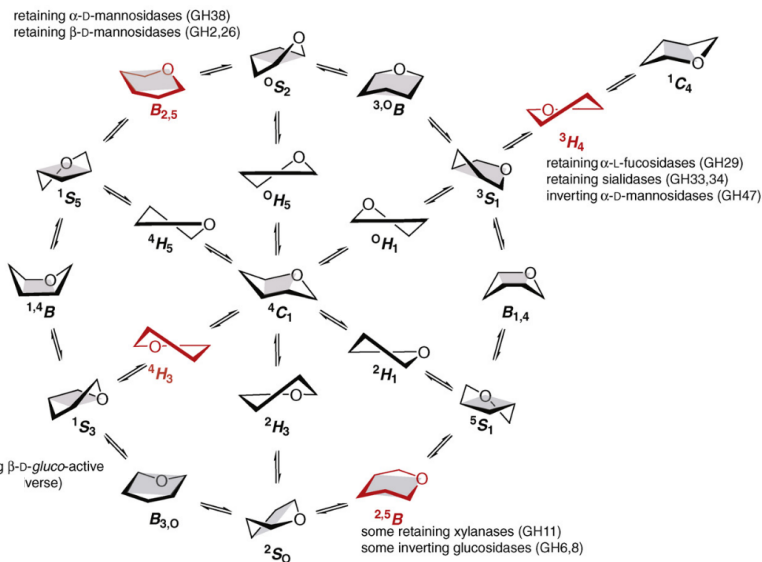


➤ Interconversion of sugar ring conformations

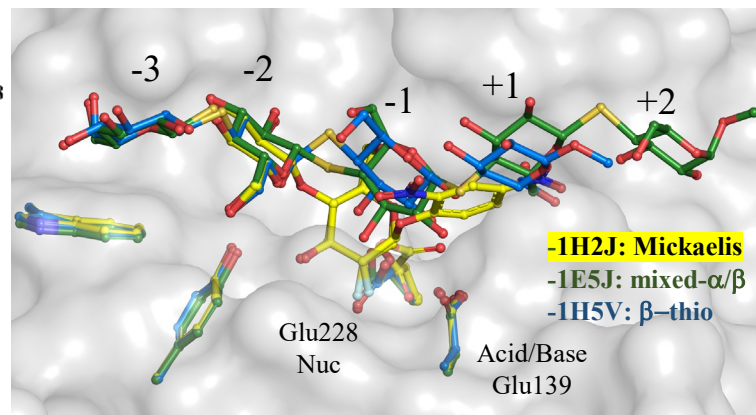
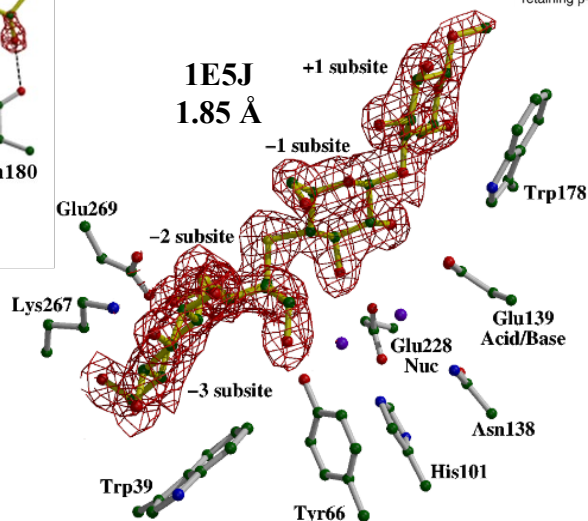
- Highlights transition state
- Essential for inhibitor design

➤ Serendipity

- New inhibitors class for glucosidases



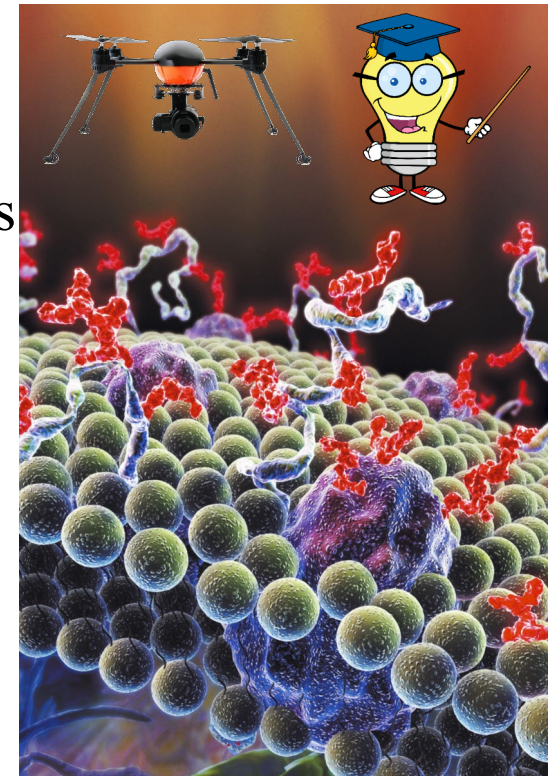
1H5V
1.1 Å



Lectins



- Ubiquitous carbohydrate binding proteins
- Specific and reversible sugar binding without modification
- Decipher the glycodecode
- Implicated in many cellular processes
 - From warning their kin to poisoning their enemies
- Multivalency compensates for low affinity
 - Give ability to agglutinate cells
- New database: <https://unilectin.unige.ch/>



Swiss Institute of Bioinformatics



UNIVERSITÉ DE GENÈVE



Unified exploration platform for manually curated and predicted lectins

Lectin structure

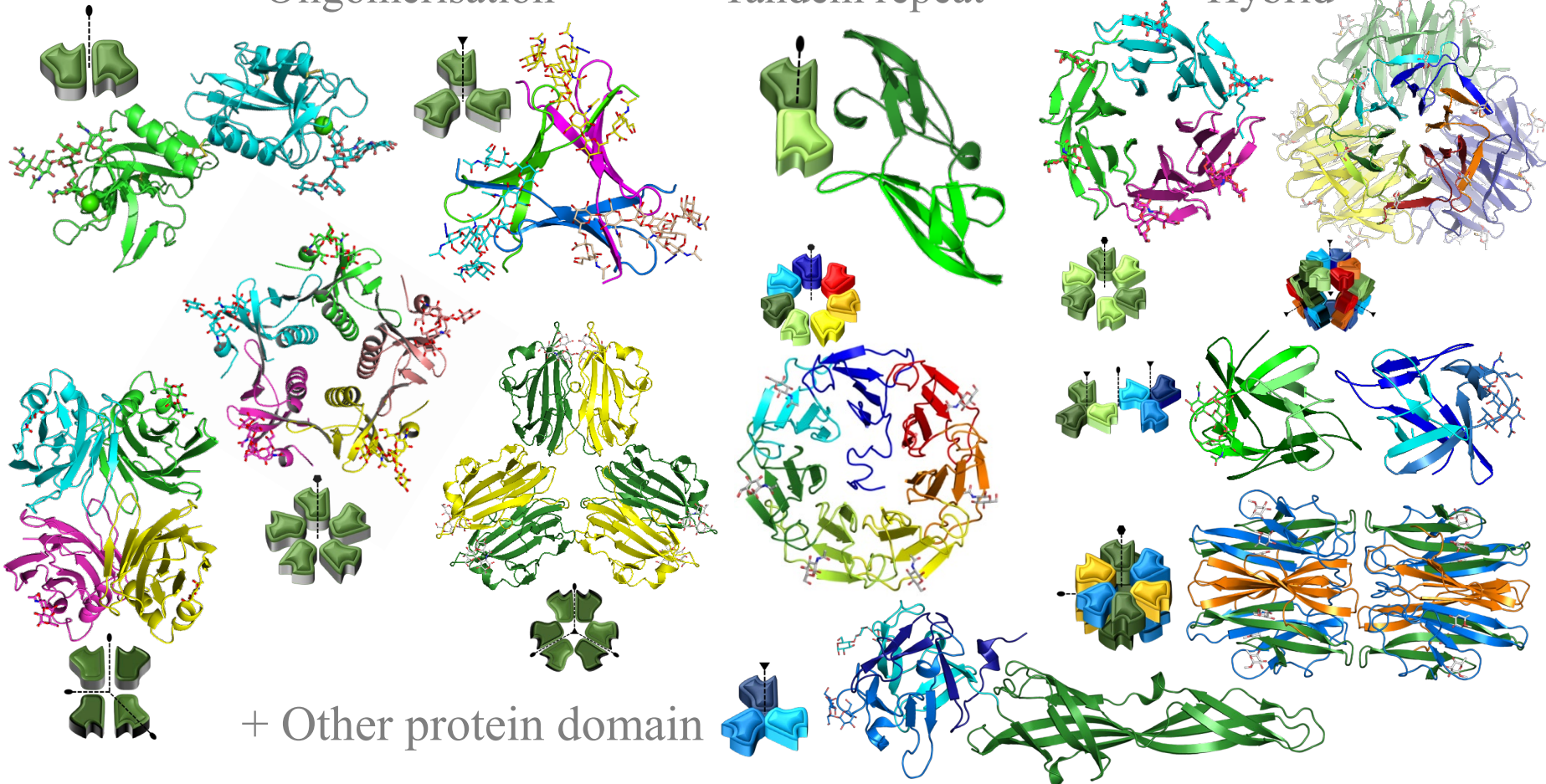


➤ High diversity of fold and quaternary structure

Oligomerisation

Tandem repeat

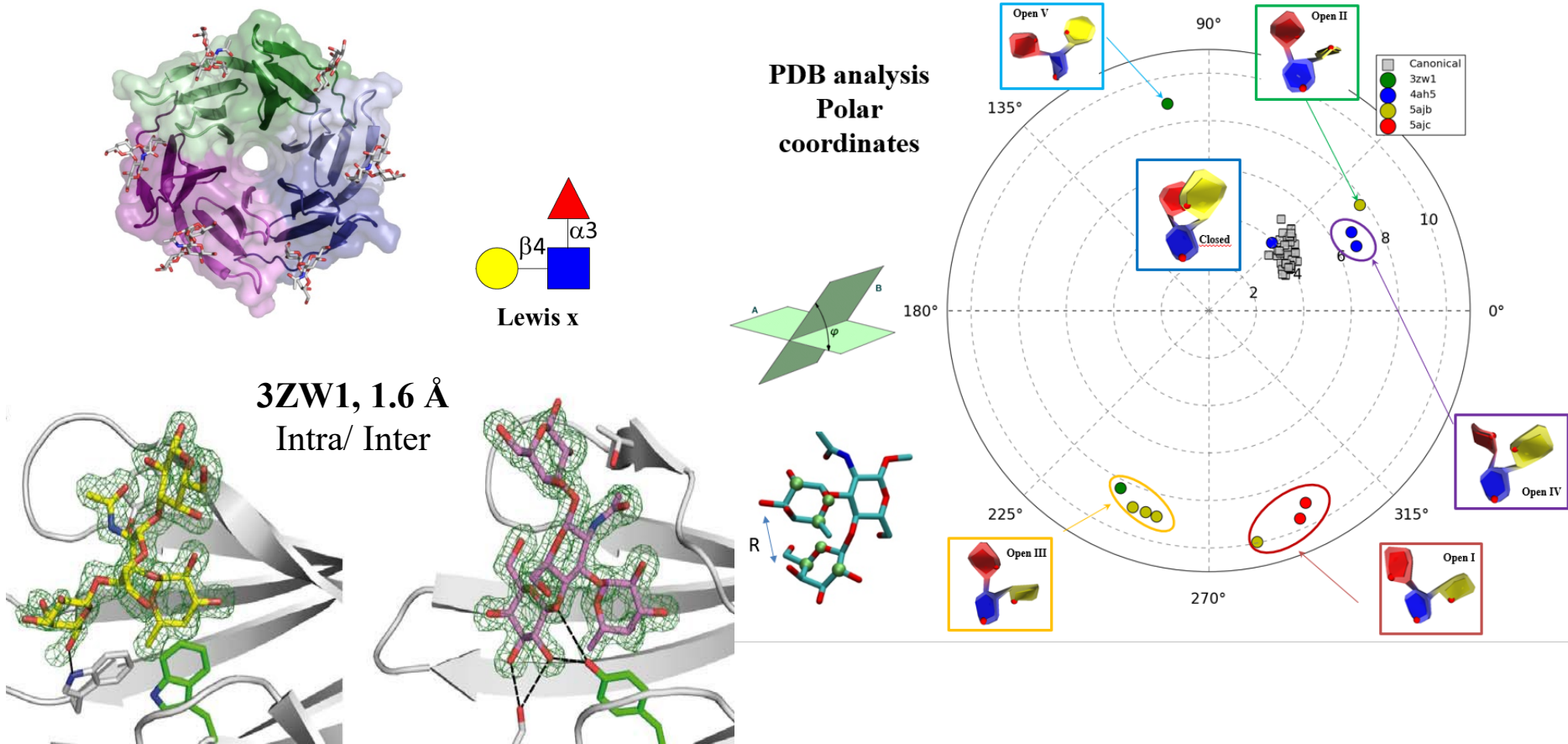
Hybrid



Rare conformation



- 1 structure of lectin complex with ring distortion
- BambL/RSL: Hidden conformation of Lewis X

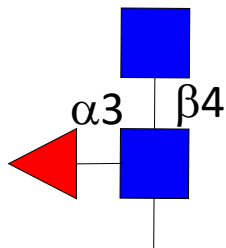


Multivalency and function



➤ CCL2 from *Coprinopsis cinerea*

- Involved in innate immunity and defense
- Toxic for nematodes and flies
 - Recognised 3-core fucose of midgut N-glycans



- Monomer on Superdex75
- RMN structure: β -trefoil fold
 - Only subsite β functional
- Could not explain toxicity
 - Monomer and monovalent

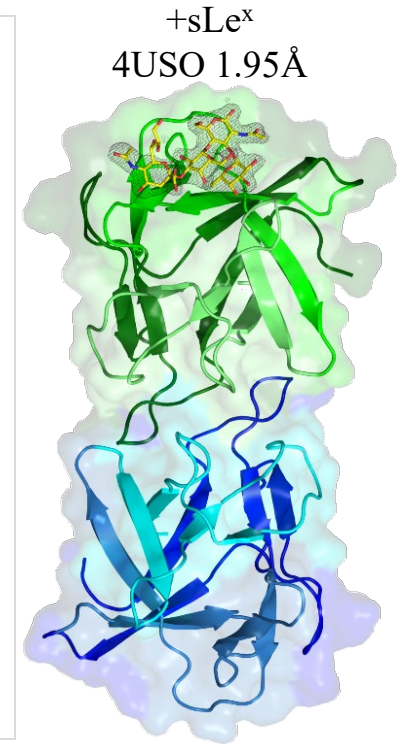
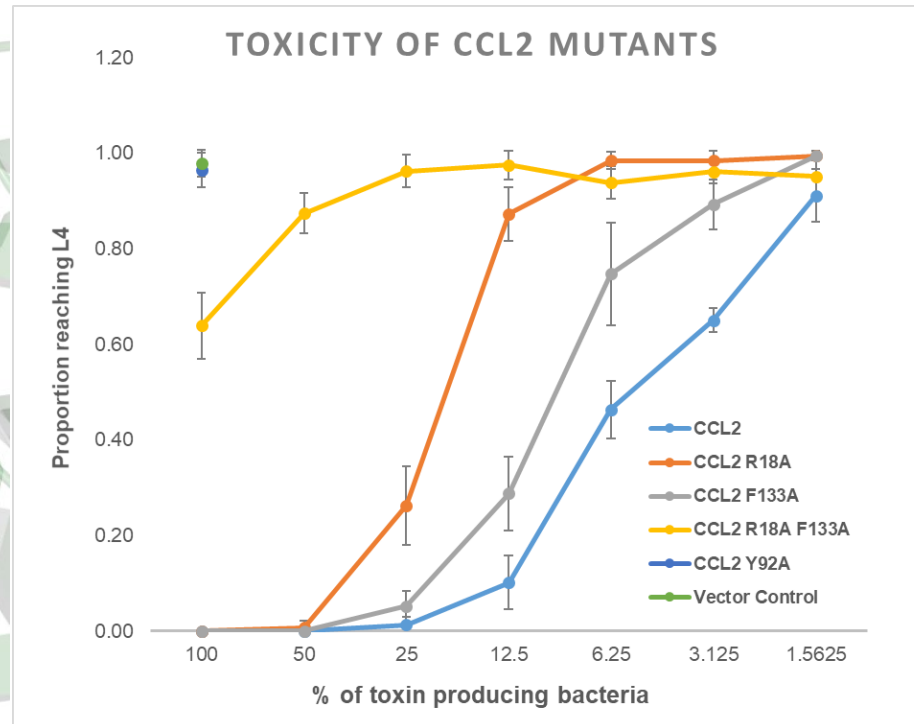
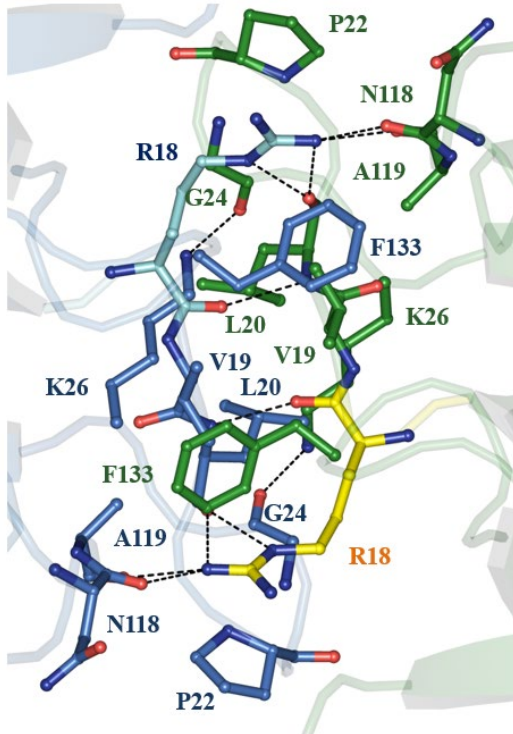
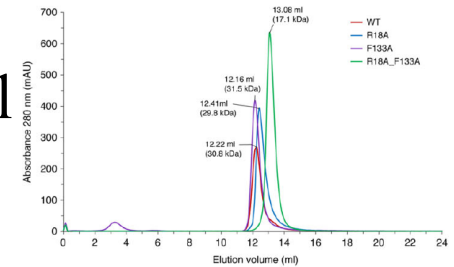


Multivalency and function-2



➤ CCL2 is a dimer

- Confirmed by SEC on Enrich70, DLS, Native gel
- Toxicity dependent on its dimerization
 - Disruption of dimer interface requires double mutation

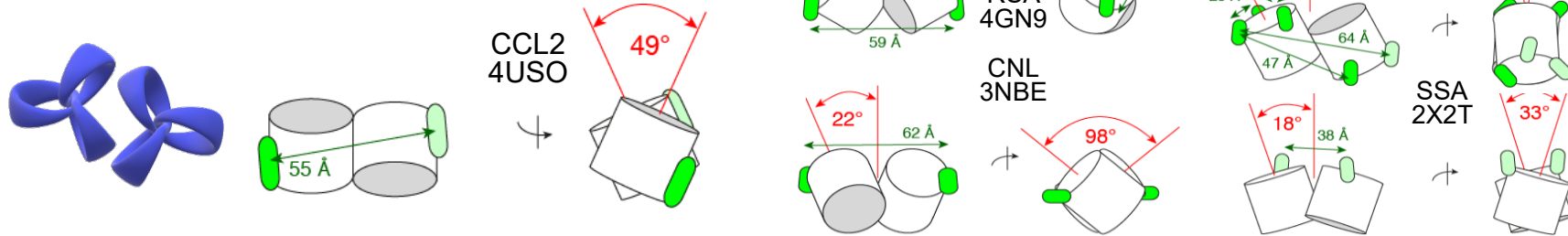


Multivalency and function-2



➤ Spatial distribution of binding sites impact function and recognition mode of β -trefoil lectins

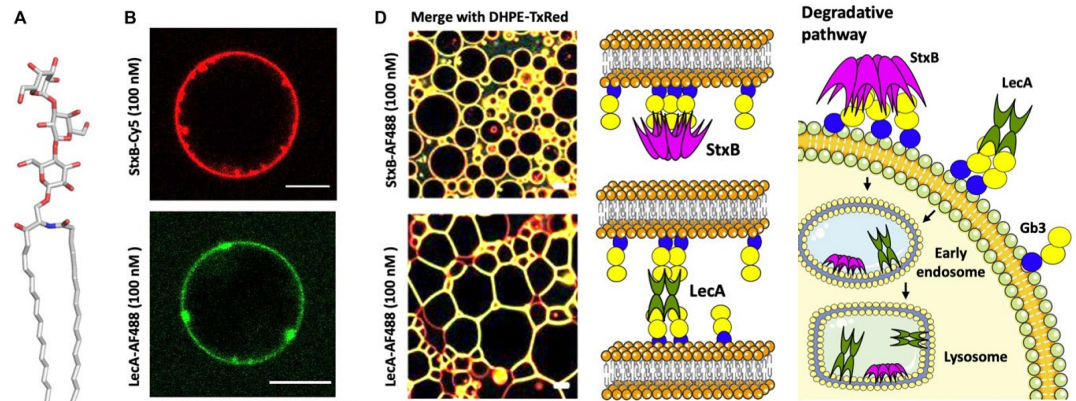
- CCL2: 1 binding surface on the side



- Others: 1/2 binding surfaces on the top

➤ Reorganisation of membrane glycoconjugates

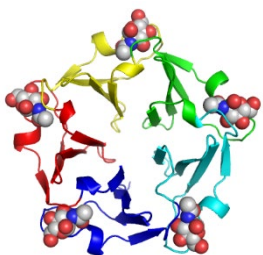
- Clustering of glycolipids
- Crosslinking
- Change in membrane dynamics
- Endocytosis



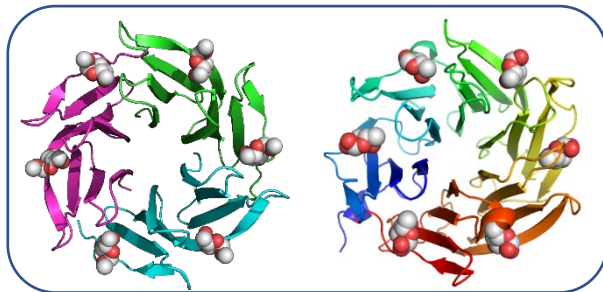
Lectins with β -propeller fold



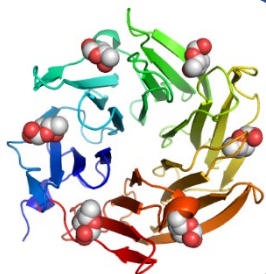
Tachylectin 2
GlcNAc



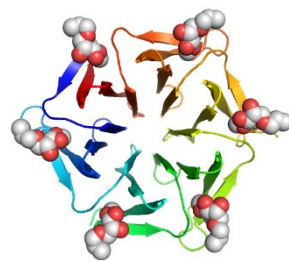
RSL
fucose



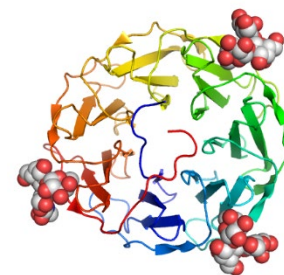
FleA
fucose



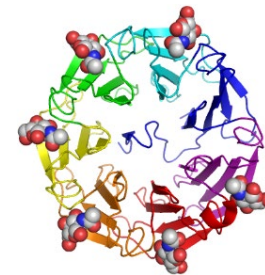
Tectonin/Lb-Tec2
Me-sugar



PLL
fucose



PVL
GlcNAc



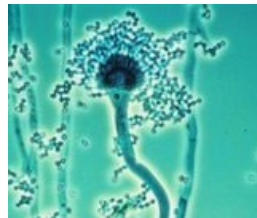
Tachipleus tridentatus

Beisel *et al.*
EMBO J. 1999



Ralstonia solanacearum

Kostlanova *et al.*
J. Biol. Chem 2005



Aspergillus fumigatus

Houser J, *Plos One*,
2013, 8:e83077



Laccaria bicolor

Sommer *et al.*
Structure 2018



Photorhabdus luminescens

Kumar *et al.*
J. Biol. Chem 2016



Pstatyrella velutina

Cioci *et al.*
J. Mol. Biol. 2006

➤ Define blade signature for prediction: PropLec in Unilectin

PropLec5A

FLF



PropLec6A

RVY



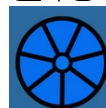
PropLec6B

GVN



PropLec7A

EVF



PropLec7B

GFG



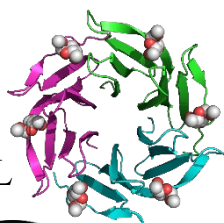
PropLec7C



Distribution of predicted blade in PropLec6A



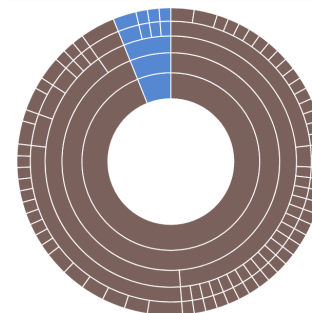
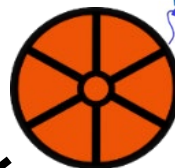
RSL
BamBL



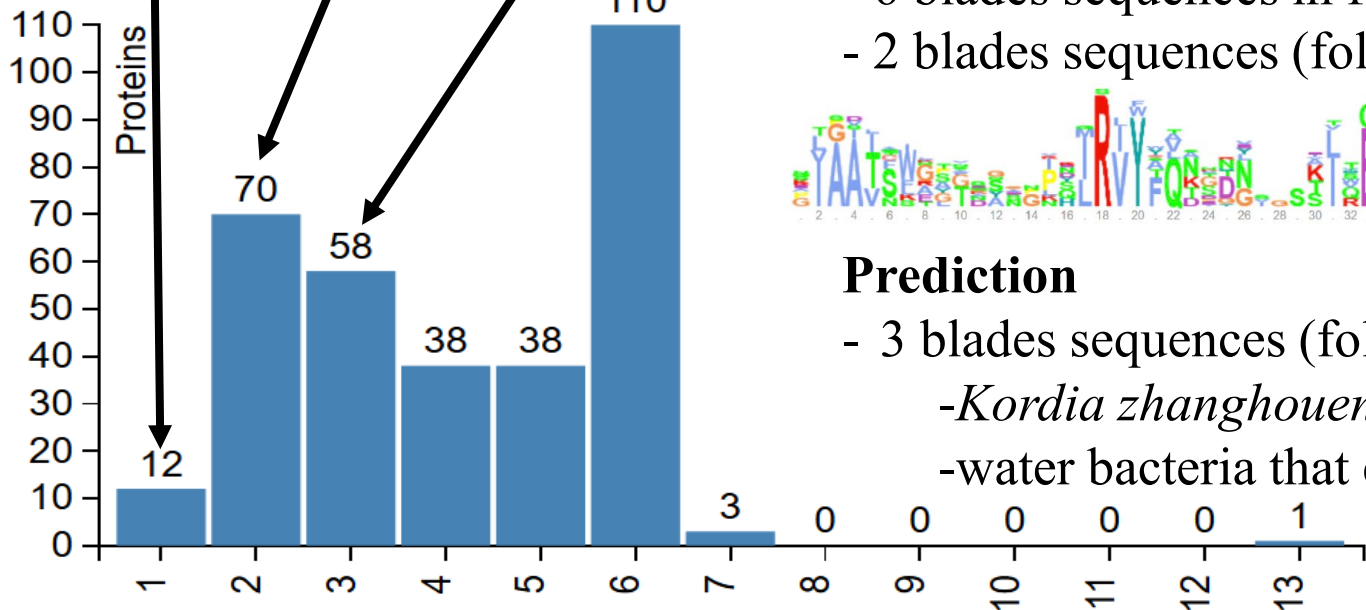
?



FleA
AAL

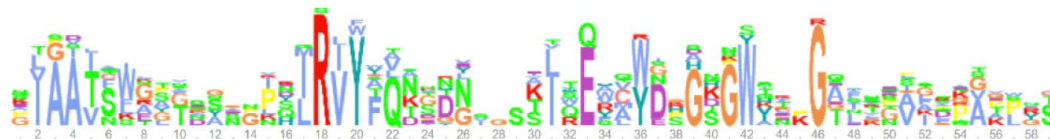


Bacteria
Eukaryota
Archaea



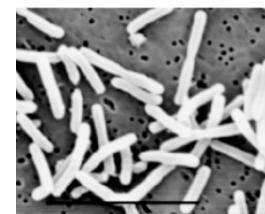
Already observed

- 6 blades sequences in fungi
- 2 blades sequences (fold : 3 x 2 blades)



Prediction

- 3 blades sequences (fold : 2 x 3 blades)
- *Kordia zhanghouensis*
- water bacteria that eats algi

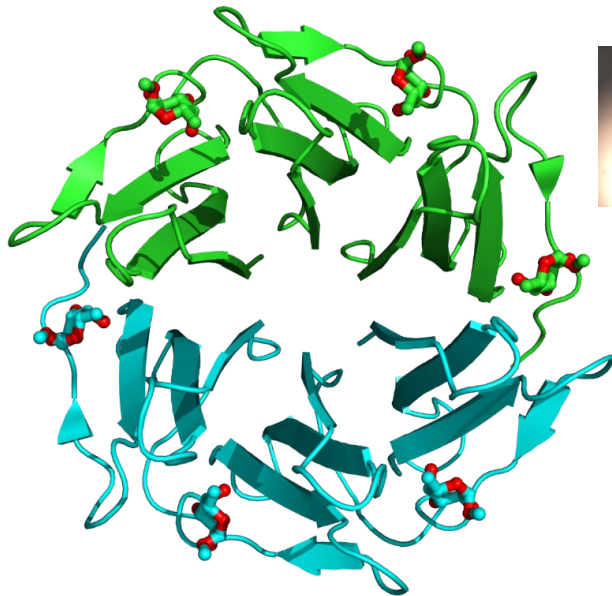
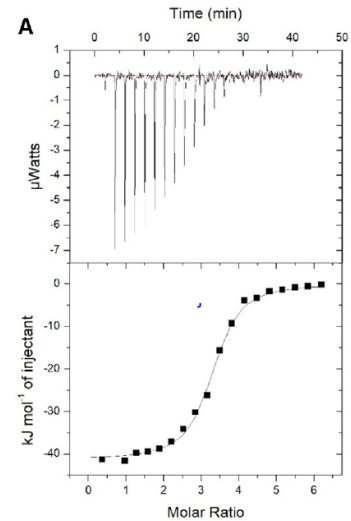


Recombinant KozL

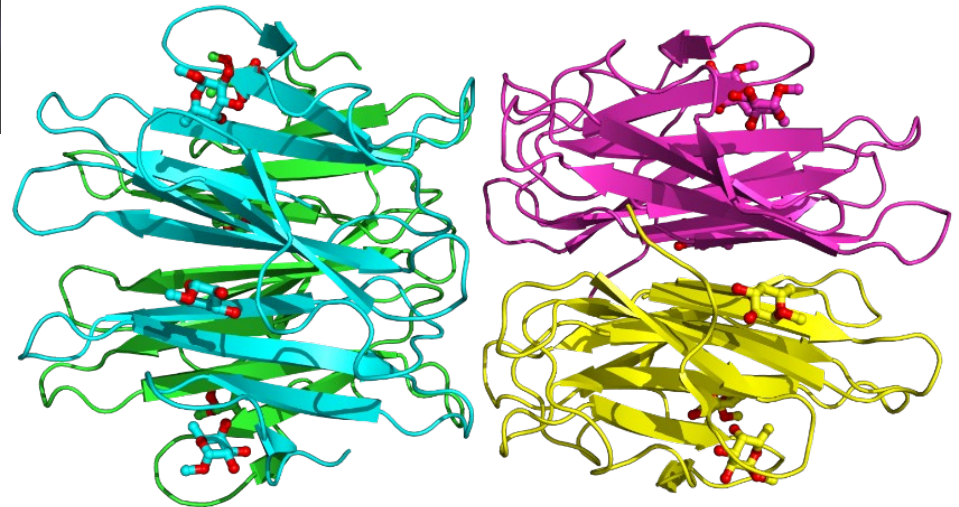


- Overexpressed in *Escherichia coli*
 - Purified on mannose agarose: 35 mg/L
 - ITC: 3 fucose binding sites
 - Tetramer by AUC

- Structure solved by SAD using α SeMeFuc
 - **Prediction corroborated: 2 x 3 blades**

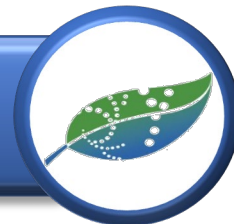


6HTN
1.55 Å

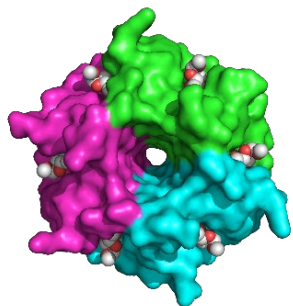




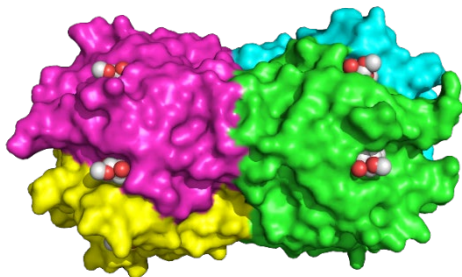
PropLec6: many arrangements



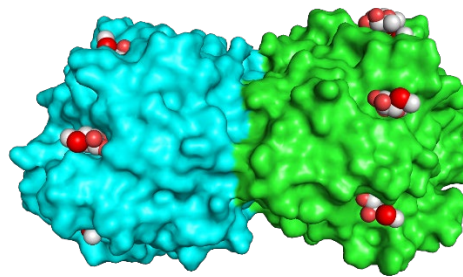
BambL/RSL
6 Fuc



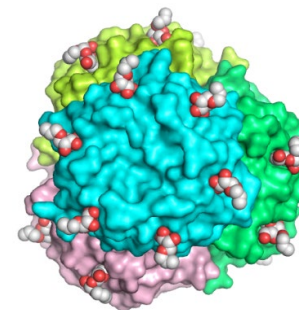
KhozL
12 Fuc



FleA/ SapL1
12 Fuc

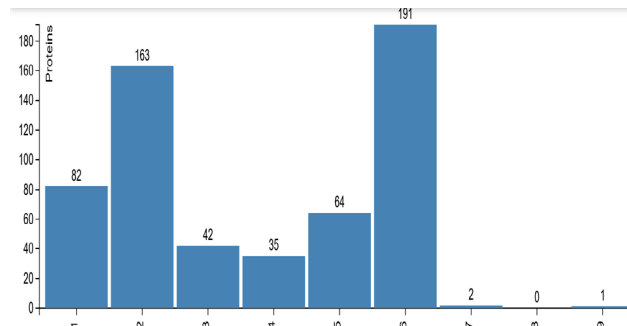


Lb-Tect2
24 Me-sug



➤ Is 1 blade occurring?

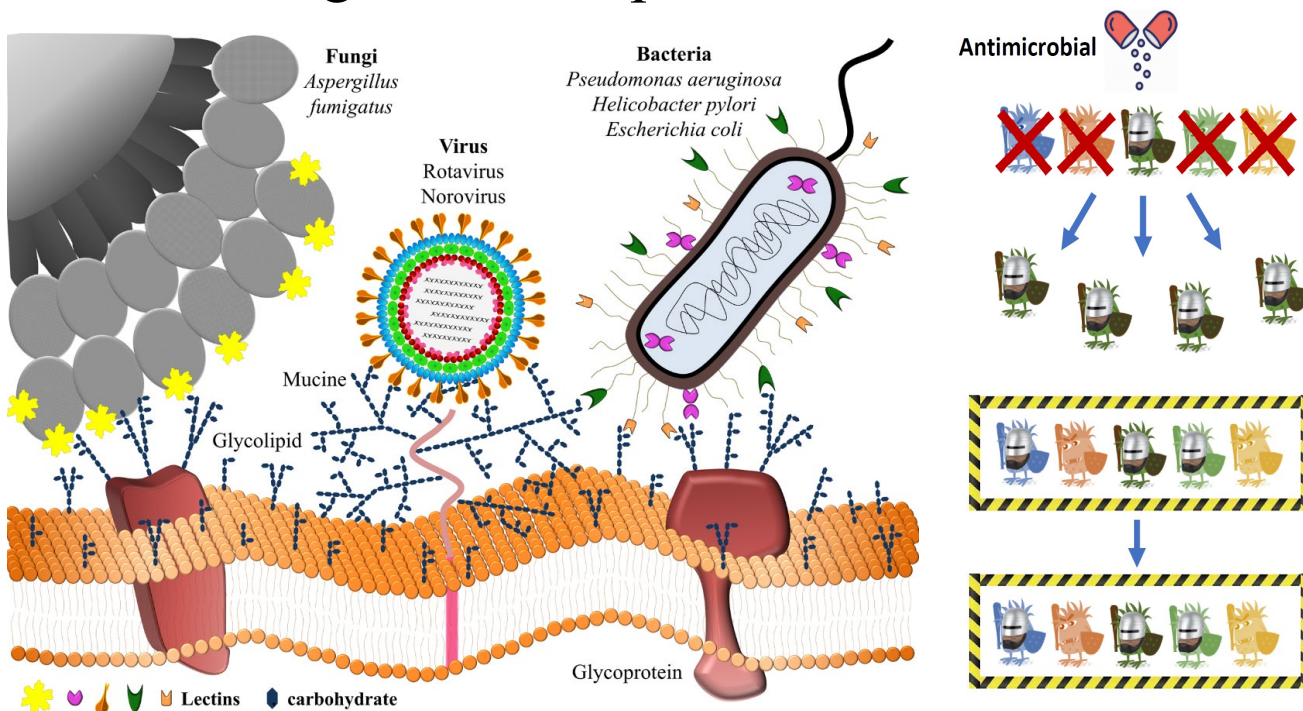
- Fold: 6x1 blade



Lectins as therapeutic targets



- Lectins used host glycoconjugates as entry points
 - Mediate host recognition and adhesion
 - Blocking lectins → prevent infections



Classic:
Destroy pathogens
→ selective pressure
driving **resistance**

vs

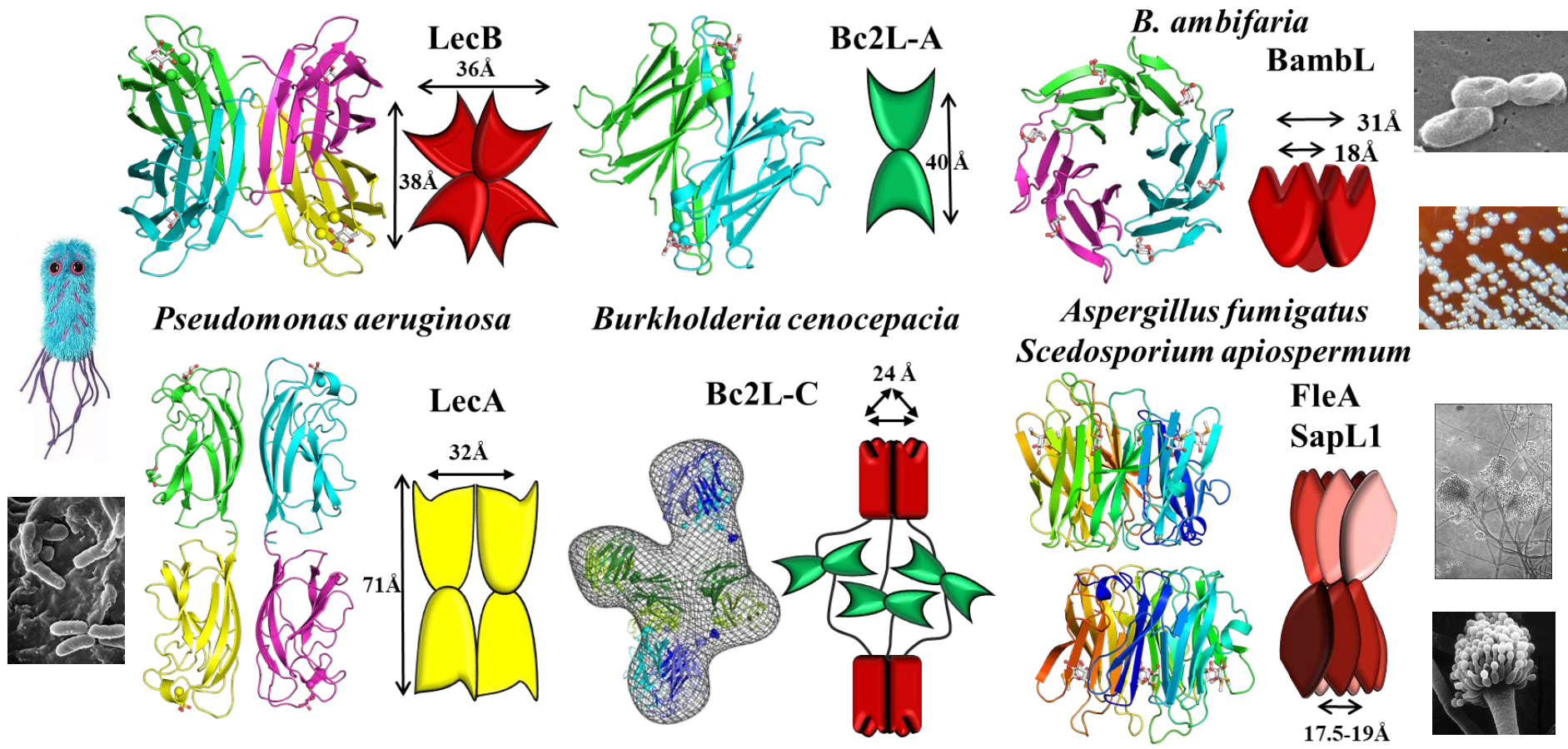
Alternative:
Disable pathogens
→ **Resistance** not
required for survival
Antiadhesion therapy

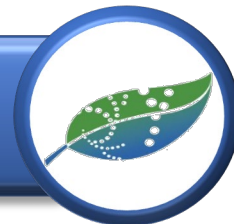
➔ **Development of glycocompounds as antimicrobials**

Lectin targets



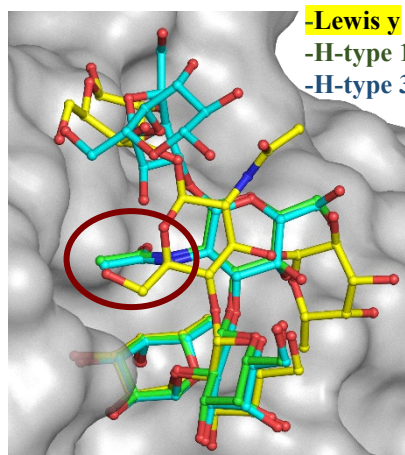
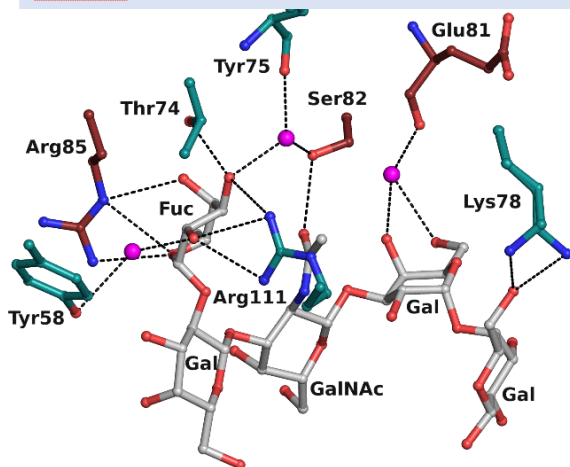
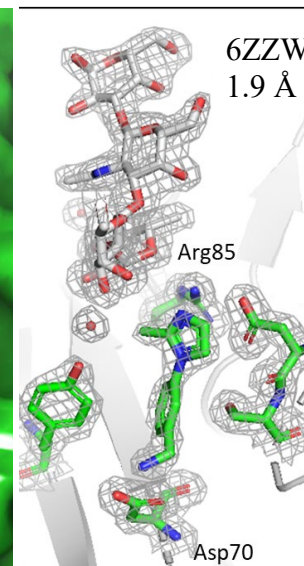
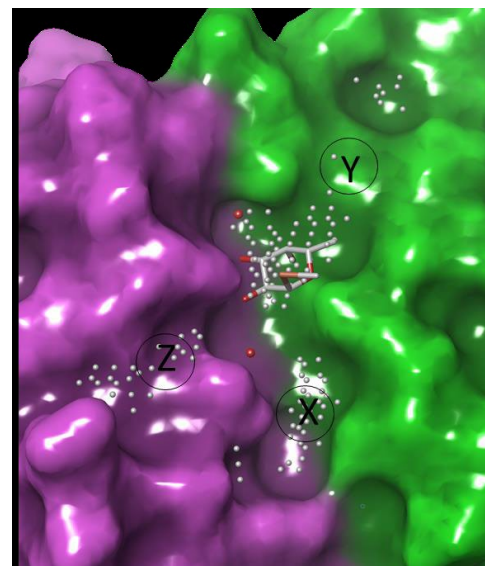
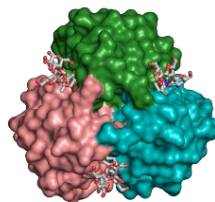
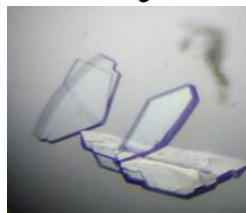
➤ Opportunistic pathogens → bronchopulmonary infections



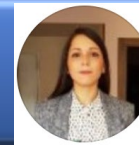
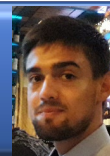


- New construct 1-132 in pCold-TEV: good yield & stable
- Complex with oligos by cocrys and with inhibitors by soaking

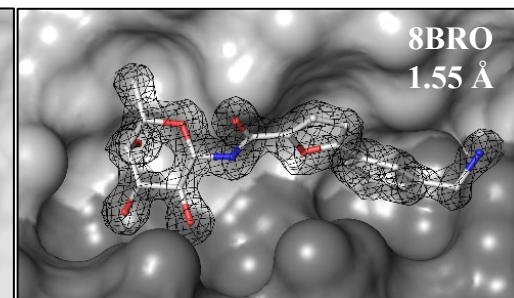
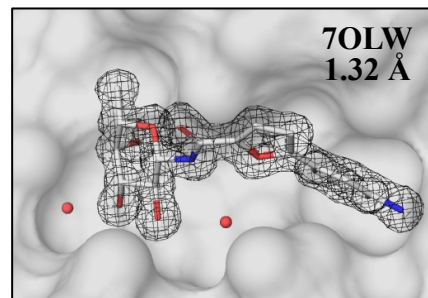
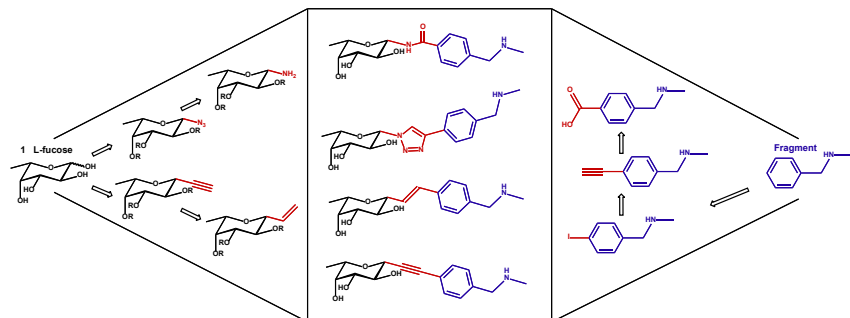
Ligand		k_D (μ M)	PDB	Res (\AA)
L-Fuc		2430	2WQ4	1.42
L-Gal		2000		
Disac		2500		
H-type 1 tri		25.4		
H-type 1 tetra		56.6	6TID	1.6
Lewis Y penta		53.2	7OLU	1.6
H-type 3-GloboH hexa		26.7	6TIG	1.9



- 3 druggable sites predicted with SiteMap in Maestro

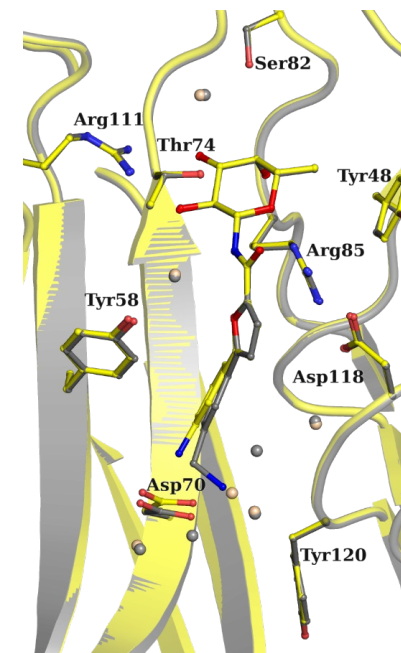
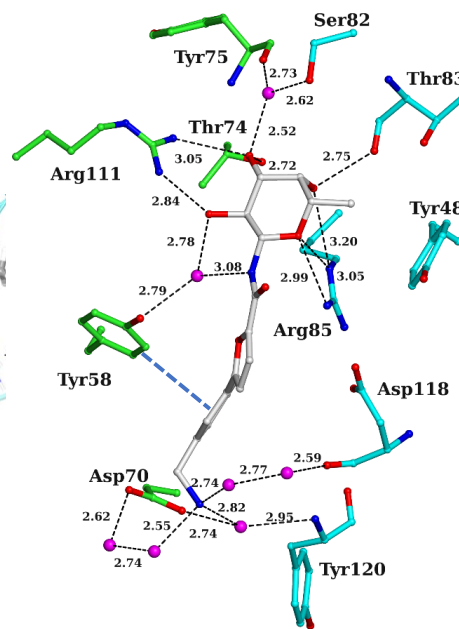
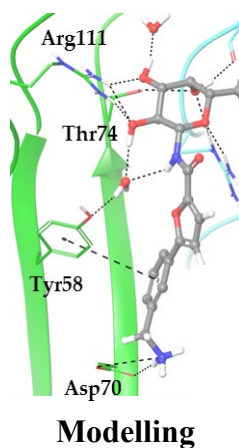


➤ Design, synthesis & evaluation of fucosides derivatives



- Transform aniline in aminomethylene
 - One order of magnitude gain
 - Terminal amine does indirect Hbonds

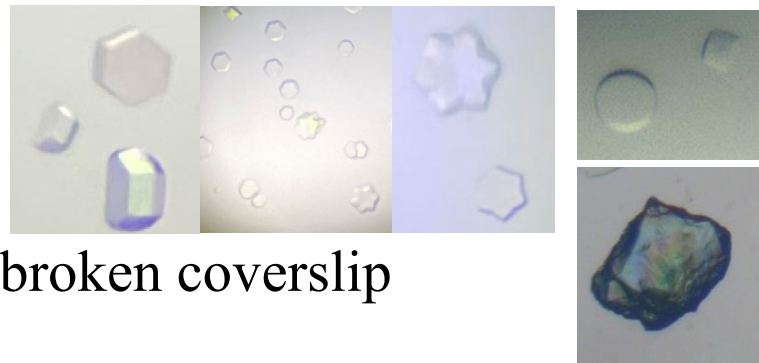
Ligand	k_D (μM)	PDB	Res (\AA)
L-Fuc αMe	2700		
22a	280	70LU	1.6
8c	nd	70LW	1.32
3	159	8BRO	1.55
4	390		





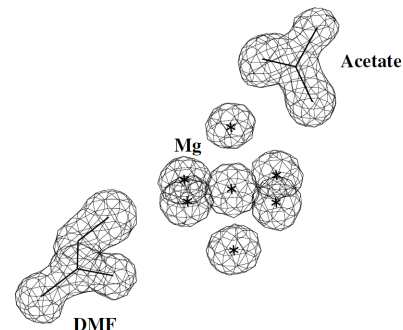
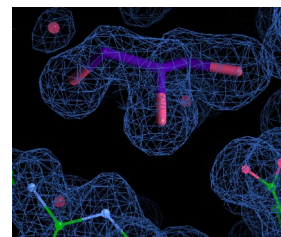
➤ Crystal shape can be misleading

- Bc21Cnt 0.9-1.2 Trisodium citrate pH 7
 - Hexagons OK - flower not OK
- Coda: 1 crystal after several month and broken coverslip
 - Not multiple, 1.3 Å



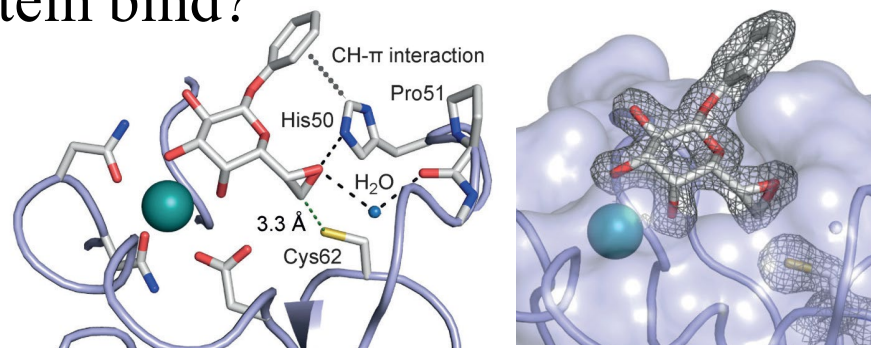
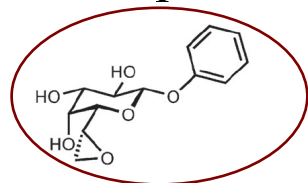
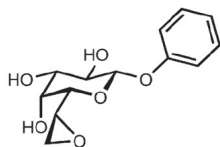
➤ Components from purification or crystallisation

- Electron density of chemical can help
 - Glycerol from cryo in TrfbL1 native
 - Cel6A D405N-SDP5: 1OC7, 1.1Å



➤ Which diastereoisomer did the protein bind?

- LecA with potential covalent epoxide
 - Crystal at unreactive pH : 4.5





Conclusions



- Still need for X-ray crystallography to gain access to protein-sugar interactions at the atomic level
 - Highest the resolution the better to draw accurate conclusions
 - Distorsion / High energy conformation
 - Real or artefact from user/program errors
 - Binding site location and architecture
 - Structure based drug design
 - Site directed mutagenesis: new specificity
 - Quaternary structure and multivalency
 - Gain fondamental knowledge on protein folding and function
 - Design new molecular tools

- Could be tricky for non glycobioologists
 - No « Structural glycobiology for dummies »
 - Do not hesitate to contact experts

Acknowledgements



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Johan Turkenburg

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Roman Sommer
Stefanie Wagner
Dirk Hauck

+++

